Ischemia-Induced Neural Stem/Progenitor Cells Within the Post-Stroke Cortex in Adult Brains

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1. Introduction

Stroke is one of the major causes of death and disability in developed countries. The central nervous system (CNS) is known for its limited reparative capacity, but several studies demonstrated that the CNS has some reparative potential and cerebral ischemia is followed by activation of endogenous neurogenesis (Nakatomi et al., 2002; Taguchi et al., 2004). It is well-known that new neurons are continuously generated in specific brain regions such as the subventricular zones (SVZ) (Alvarez-Buylla et al., 2002) and the subgranular zone within the dentate gyrus of the hippocampus (SGZ) (Kuhn et al., 1996). Although adult cerebral cortical neurogenesis remains controversial, accumulating evidence has shown that under pathological conditions, new neurons are generated in the adult mammalian cerebral cortex (Magavi et al., 2000; Jiang et al., 2001; Jin et al., 2006; Yang et al., 2007). This suggests that neural stem/progenitor cells (NSPCs) can be activated in the cortex by brain injury such as ischemic stroke. In support of this notion, we demonstrated that NSPCs develop in the post-stroke area of the cortex in the adult murine (Nakagomi et al., 2009a; Nakagomi et al., 2009b; Nakano-Doi et al., 2010; Saino et al., 2010) and human brain (Nakayama et al., 2010), and we referred to these as ischemia/injury-induced NSPCs (iNSPCs). These cells express markers of NSPCs, such as nestin and Sox2. They also form neurospheres that have the capacity for self-renewal, and differentiate into electrophysiologically functional neurons, astrocytes, and myelin-producing oligodendrocytes (Nakagomi et al., 2009a; Nakagomi et al., 2009b; Nakano-Doi et al., 2010; Clausen et al., 2011). In addition, we demonstrated that iNSPCs originate, at least in part, from within the cerebral cortex, but not from SVZ cells (Nakagomi et al., 2009b). However, the detailed origin and identity of the iNSPCs remains unclear. In this chapter, we introduce the characterization and possible origin of iNSPCs based on our reports and recent viewpoint, and compare them to other previously reported types of CNS stem/progenitor cells, including SVZ astrocytes (Doetsch et al., 1999), ependymal cells (Moreno-Manzano et al., 2009), reactive astrocytes (Shimada et al., 2010), resident glia (Zawadzka et al., 2010), and oligodendrocyte precursor cells (OPCs) (Kondo et al., 2000). We also refer to the possible cortical neurogenesis by iNSPCs and to the therapeutic potential of iNSPC transplantation in stroke patients.
2. NSPCs in the adult cortex

In the CNS of adult mammals, it is well-known that NSPCs are present in the SVZ and SVG, and that ongoing neurogenesis is retained in these two zones. However, accumulating evidence suggests that NSPCs reside in many parts of the adult brain including the cortex (Arsenijevic et al., 2001; Joh et al., 2005; Kallur et al., 2006; Jiao et al., 2008; Willaime-Morawek et al., 2008), striatum (Kallur et al., 2006; Willaime-Morawek et al., 2008), subcortical white matter (Nunes et al., 2003), and spinal cord (Weiss et al., 1996; Parr et al., 2008). These observations suggest that NSPCs are widely distributed throughout the adult CNS. In this chapter, we introduce iNSPCs, which are induced within the post-stroke cortex after brain injury/ischemia in adult brains.

2.1 Cortical development in the embryonic stage: comparison to iNSPCs in the cortex

In the embryonic stage, neurogenesis was observed throughout the CNS including the cortex. Mignone and colleagues traced nestin-expressing NSPCs, and showed that green fluorescent protein (GFP) expression in developing transgenic nestin-GFP mice was evident on as early as day 7 of embryonic development (e7). At e8, a GFP signal was observed predominantly in the neural plate, and by e10 intense GFP fluorescence was observed throughout the neuroepithelium. At e10 to e12, GFP signals marked the entire thickness of the cerebral wall, but GFP expression became weaker near the pial surface and stronger in the ventricular zones starting from e12. Finally, in the adult brain, GFP was selectively expressed in the SVZ and SGZ in areas related to continuous neurogenesis (Mignone et al., 2004). Thus, in the postnatal CNS, constitutive neurogenesis is known to be retained in only two regions the SVZ (Alvarez-Buylla et al., 2002) and SGZ (Kuhn et al., 1996). However, under pathological conditions, neurogenesis may occur again in the adult cerebral cortex (Magavi et al., 2000; Jiang et al., 2001; Jin et al., 2006; Yang et al., 2007). Supporting their observations, nestin-positive NSPCs were observed after brain injury/ischemia in nonconventional neurogenic zones, such as the cortex (Nakagomi et al., 2009b; Nakayama et al., 2010). Because they were rarely observed in the absence of brain injury (Nakagomi et al., 2009b), cortical neurogenesis may reoccur only in the case of brain injury. These findings suggest that in adult mammalian brains, NSPC activation and neuronal homeostasis are maintained under physiological conditions, at least in part, in specific brain regions, such as the SVZ and SGZ. However, after brain injury, it appears that regional NSPCs are mobilized to accelerate tissue repair by a mechanism similar to embryonic neurogenesis. Taken together, these observations suggest that ischemia/hypoxia is essential for the induction of NSPCs in the adult cortex, although we remain unaware of the required signaling and/or factors.

2.1.1 Characteristics of iNSPCs from the post-stroke cortex

To confirm the possible adult neurogenesis induced by brain injury, we have sought to isolate NSPCs from the injured area of the post-stroke cortex. Previously, we established a highly reproducible murine model of cortical infarction using CB-17/Icr-+/+Jcl and CB-17/Icr-Scid/scid Jcl mice. The infarct area in mice of this background has been limited to the ipsilateral cerebral cortex of the territory occupied by the middle cerebral artery (MCA) (Taguchi et al., 2004; Taguchi et al., 2007; Nakagomi et al., 2009a; Nakagomi et al., 2009b;
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Nakano-Doi et al., 2010; Saino et al., 2010; Taguchi et al., 2010). Following MCA occlusion, abundant nestin-positive cells emerged within the post-stroke cortex, although they were rarely observed in the non-ischemic cortex (Nakagomi et al., 2009b; Nakano-Doi et al., 2010; Saino et al., 2010). To examine whether these cells showed stem cell-characteristics, we cultured cells isolated from the post-stroke cortex under conditions that promoted the formation of neurospheres (Reynolds et al., 1992). In brief, tissue from the ischemic core of the post-infarct cerebral cortex was obtained on day 7 after MCA occlusion. Cells were dissociated by passage through 23 and 27 gauge needles, and cell suspensions were incubated in tissue culture flasks with DMEM containing epidermal growth factor (EGF), basic fibroblast growth factor (bFGF) and N2 supplement. This procedure allowed us to obtain nestin-positive neurosphere-like cell clusters (iNSPCs) (Nakagomi et al., 2009a; Nakagomi et al., 2009b; Nakano-Doi et al., 2010) (Fig. 1).

![Fig. 1. Isolation of nestin-positive iNSPCs developing within the post-stroke cortex](image)

However, iNSPCs were rarely obtained in the absence of brain injury. Notably, because we could not obtain iNSPCs from the peri-stroke cortex, it is possible that these cells are generated within the degenerating cortical tissue after ischemic stroke.

Uptake of 5-bromo-2′-deoxyuridine (BrdU) by iNSPCs was confirmed in vivo (Nakano-Doi et al., 2010; Saino et al., 2010) and in vitro (Nakagomi et al., 2009a; Nakagomi et al., 2009b), showing that they have the proliferative activity. They possessed self-renewal capacity, which was confirmed by a clonal assay. However, in contrast to the embryonic stem cells, the cluster formation in the same medium at a clonal density was limited to between three and five cell passages (Nakagomi et al., 2009b; Nakano-Doi et al., 2010), consistent with other adult candidates of stem cells, such as neurospheres derived from hippocampus (Bull et al., 2005) and subcortical white matter (Nunes et al., 2003). These observations suggest that cortex-derived iNSPCs are more likely to be neural progenitors than neural stem cells (NSCs). However, they certainly differentiated into electrophysiologically functional...
neurons, astrocytes, and myelin-producing oligodendrocytes (Nakagomi et al., 2009a; Nakagomi et al., 2009b; Nakano-Doi et al., 2010), indicating that iNSPCs have a stemness-capacity similar to other adult NSPCs. Interestingly, they predominantly differentiated into neurons (approximately 35%) and oligodendrocytes (approximately 30%) rather than astrocytes (approximately 5%) (Nakagomi et al., 2009a; Nakano-Doi et al., 2010; Nakagomi et al., 2011) with characteristics discriminating from other adult NSPCs such as SVZ astrocytes, most of which are known to differentiate into astrocytes. These findings suggest that iNSPCs have a strong potential of contributing to cortical neurogenesis compared to NSPCs derived from other origins, especially under the conditions of brain injury.

Consistent with the SVZ-derived NSPCs (Kim et al., 2009b), cortical iNSPCs expressed several pluripotent/undifferentiated cell markers, including Sox2, Klf4, c-myc and Nanog (Nakagomi et al., 2009b; Nakagomi et al., 2011). However, expression of various pluripotent/undifferentiated cell markers was not observed in the cortex without brain injury. These observations suggest that cell reprogramming may occur in unknown cells of the cortex in response to brain injury/ischemia, thereby promoting the induction of iNSPCs. However, further studies are needed to clarify this hypothesis.

2.1.2 Comparison to other types of reported CNS stem/progenitor cells

Accumulating evidence has shown several candidates for adult NSPCs, which can contribute to adult neurogenesis in the cerebral cortex. One of these candidates may be radial glia cells, which are derived from neuroepithelial cells and functions as NSPCs during development. The radial glia are able to develop into several types of NSPCs, such as SVZ astrocytes, ependymal cells, and OPCs in adult (Kriegstein et al., 2009). However, precise cell source of cortical NSPCs remains unclear, especially in the injured brain.

Previous studies demonstrated that SVZ astrocytes have the capacity to migrate towards injured lesions, including the cerebral cortex (Goings et al., 2004). However, our study using GFP-expressing vector, failed to demonstrate cell migration from the SVZ to the cortex after cerebral infarction in vivo, but demonstrated that iNSPCs in the post-stroke cortex originated, at least in part, from the cerebral cortex (Nakagomi et al., 2009b). Consistently, subsequent studies showed that NSPCs developing within and around the post-stroke cortex are derived from locally activated stem/progenitor cells, but not from SVZ cells (Ohira et al., 2010; Shimada et al., 2010). To answer which cells can be activated by cerebral injury, some studies proposed the reactive astrocytes as a source of injury-induced NSPCs (Oki et al., 2010; Shimada et al., 2010), because NSPCs express the astrocyte marker, GFAP. However, we could not detect GFAP- (Nakagomi et al., 2011) and S100β-positive astrocytes within the post-stroke cortex (Nakagomi et al., 2009b). Eventually, the isolated iNSPCs from the infarct cortex rarely expressed GFAP and developed few astrocytic traits even after differentiation (Nakagomi et al., 2011). In addition, although we found some nestin and GFAP double-positive reactive astrocyte-like cells in the peri-infarct area, we could not obtain neurospheres from these areas. These findings strongly suggest that the source of iNSPCs within the infarct cortex is distinct from reactive astrocytes.

Currently, it is still highly controversial whether periventricular NSPCs can be derived from SVZ astrocytes, ependymal cells, or both (Chojnacki et al., 2009). Ependymal cells were originally considered to be the resident stem cell population in the wall of the lateral
ventricle, in which they locate nearby perivascular cells (Pfenninger et al., 2007; Coskun et al., 2008). Although it is controversial whether ependymal cells have NSPC activity or not, recent studies confirmed that ependymal cells do not play a role in adult neurogenesis under normal conditions, but do possess NSPC activity and can differentiate into neurons, astrocytes, and oligodendrocytes in response to the CNS injuries including ischemic stroke (Carlen et al., 2009; Moreno-Manzano et al., 2009). Furthermore, ependymal cells express PDGFRα (Danilov et al., 2009) and NG2 (Moreno-Manzano et al., 2009), and have the structure of lipid droplets, microvilli, and cilia (Coskun et al., 2008; Danilov et al., 2009). Consistent with the traits of ependymal cells, iNSPCs express PDGFRα and NG2, but do not possess microvilli-like structures (Nakagomi et al., 2011). These findings indicate that iNSPCs do not have completely identical characteristics to those of ependymal cells.

Adult OPCs comprise approximately 5%–8% of the glial cell population in the CNS. Their function in the CNS remains unknown, although accumulating evidence has shown that they have NSPC activity (Kondo et al., 2000; Gaughwin et al., 2006), in addition to myelin-producing abilities (Sundberg et al., 2010). OPCs are known to express NG2 (Ulrich et al., 2008) and PDGFRα (Hall et al., 1996), and OPCs expressing A2B5 have NSPC activity (Kondo et al., 2000; Gaughwin et al., 2006). To investigate whether iNSPCs are derived from OPCs, we analyzed OPC markers expressed by iNSPCs in vivo and in vitro. Although iNSPCs express some OPC markers such as NG2 and PDGFRα, they do not possess A2B5 or even Olig2 (another OPC marker) (Billon et al., 2002). These observations indicate that iNSPCs are different from reported multipotent OPCs (Kondo et al., 2000; Gaughwin et al., 2006). However, when iNSPCs were incubated in OPC-promoting medium (Chen et al., 2007), they began to express Olig2. In addition, almost all cells developed from iNSPCs in this medium differentiated into O4- and/or myelin-associated glycoprotein (MAG)-positive oligodendrocytes (Nakagomi et al., 2011). These findings suggest that iNSPCs express some OPC markers during their development/differentiation.

It is well-known that NG2 is not only the marker of OPC, but is also the marker of resident glial cells/glial progenitors (Stallcup et al., 1987). More recently, NG2-positive resident glia was reported to develop NSPC activity after brain injury (Yokoyama et al., 2006; Zawadzka et al., 2010). We demonstrated that cortical iNSPCs express NG2 and PDGFRα in a similar manner to resident glial/progenitor cells. However, neuronal differentiation from NG2- and/or PDGFRα-positive glial cells is rarely observed (Zawadzka et al., 2010; Richardson et al., 2011), suggesting that iNSPCs may be different from these glial cells or belonging to unknown cell type, which expresses some glial markers.

2.1.3 What is the origin of iNSPCs in the cortex?

So far, it seems possible that iNSPCs are different from previously proposed CNS stem/progenitor cells such as SVZ astrocytes, reactive astrocytes, ependymal cells, or OPCs. The essential difference of these cells may be their induction pattern and localization, because iNSPCs were found only after ischemic insult, and in close association with the blood vessels in the cortex. This unique localization allowed us to examine the characteristics of cells nearby blood vessels as a candidate of iNSPCs.

Our studies showed that the nestin-positive iNSPCs developed in the perivascular regions of the post-stroke cortex (Nakano-Doi et al., 2010; Nakayama et al., 2010), where nestin-positive
cells express NG2 and PDGFRβ (both of which are the pericyte marker), suggesting that the iNSPCs are derived from pericytes. Pericytes with multipotent progenitor activity have been indentified in various organs (Crisan et al., 2009) as well as in the CNS (Dore-Duffy et al., 2006). In addition, Dore-Duffy and colleagues (Dore-Duffy et al., 2006) showed that pericyte-derived NSPCs can be isolated from the CNS of non-injured animals. However, we hardly obtained iNSPCs from the nonischemic cortex (Nakagomi et al., 2009b), suggesting that pericytes in the cortical tissues increase their stemness activity during the progression of cerebral injury.

Increasing evidence has shown that ischemic insult promotes stem cell activity, and NSPCs (Sirko et al., 2009; Xue et al., 2009) and neuronal progenitors (Ohira et al., 2010) are also induced in response to cortical ischemic injury. These cortical NSPCs are frequently observed at the subpial/cortical layer 1 regions, suggesting that NSPCs can be activated preferentially in the cortical surface. Independent of these studies, we found nestin/Sox2-positive iNSPCs proliferating in the pia mater, which covers the surface of the post-ischemic cortex (Nakagomi et al., 2011). Pia mater is widely distributed throughout the CNS, and is closely associated with the blood vessels. It has been reported that leptomeninges (including pia mater and arachnoid membrane) regulate NSPCs (Sockanathan et al., 2009) and cortical neuron generation (Siegenthaler et al., 2009) in embryonic cortical formation, and function as a niche for stem/progenitor cells with neuronal differentiation potential (Bifari et al., 2009). These findings suggest that pia mater contains NSPCs at embryonic stage. The pial iNSPCs, which we found in the adult brain, partially spread into the cortical parenchyma as perivascular cells/pericytes with expression of pericyte markers such as NG2 and PDGFRβ. In addition, cells isolated from the infarcted area including pia mater and cortex and sorted by magnetic cell sorting (MACS) with a pericyte marker (PDGFRβ) had NSPC activity and differentiated into neurons (Nakagomi et al., 2011). These findings indicate that the microvascular pericytes that distribute from the pia mater to the cortex are a potential source of the iNSPCs (Fig. 2).

![Fig. 2. Schematic representation for the fate of iNSPCs following cortical infarction](https://www.intechopen.com)
Thus, our recent study suggests that pia mater may have the potential to generate NSPCs even in the adult brain. Until now, it has been demonstrated that pia mater, as well as some CNS pericytes, originate from the neural crest (Morse et al., 1984; Etchevers et al., 2001). We recently demonstrated that pial iNSPCs express various neural crest markers, such as Sox9, Sox10, Snail, Slug, and Twist (Aihara et al., 2010; Nakagomi et al., 2011), suggesting that pial iNSPCs are neural crest derivatives. This may provide a novel concept that neural crest-derived cells play a crucial role in the CNS repair following cortical infarction by a similar mechanism to the CNS formation in development. Considering that the neural crest has stem cell potential (neural crest-derived stem cells) (Teng et al., 2006) with differentiation into a variety of cell types including neurons and glia (Nagoshi et al., 2009), this hypothesis would not be surprising. In addition, this might explain why Schwann cells, which have neural crest origin, are induced in the injured CNS (Zawadzka et al., 2010). However, the precise source, lineage, and traits of iNSPCs warrant further investigation. This may be clarified through experiment of lineage labeling by genetic means.

2.2 Potential contribution of endogenous iNSPCs to cortical neurogenesis

Although iNSPCs are generated within the post-stroke area following cortical infarction, almost all of them can undergo apoptotic cell death (Saino et al., 2010). Subsequently, appropriate support for survival of iNSPCs is essential in maintaining post-stroke neurogenesis. Because iNSPCs developed in close association with the blood vessels from the pia mater to the cortex, they must be influenced by the vascular microenvironment, consisting of endothelial cells (ECs) (Palmer et al., 2000; Louissaint et al., 2002) and inflammatory cells infiltrated after cerebral injury (Saino et al., 2010).

ECs are a component of the blood brain barrier (BBB) and also function as a vascular niche (Shen et al., 2008). It has been reported that although inflammation exacerbates post-stroke neuronal damage, inflammation is a strong stimulus for activation of neurogenesis. Such inflammatory reactions may happen in perivascular (Virchow-Robin) spaces (Hutchings et al., 1986), in which inflammatory cells such as macrophage and lymphocytes infiltrate and may affect angiogenesis and neurogenesis after brain injury. These factors should be considered when observing cortical neurogenesis through iNSPCs after ischemic stroke.

NSPCs reside in a vascular niche and the vasculature is regarded as a key element, especially in the adult SVZ (Tavazoie et al., 2008). ECs are believed to make valuable contribution to this vascular microenvironment (Palmer et al., 2000; Louissaint et al., 2002). In support of this viewpoint, co-culture experiments showed that ECs increase proliferation of NSPCs derived from the adult SVZ (Shen et al., 2004; Teng et al., 2008). Furthermore, we demonstrated both in vitro and in vivo, that the presence of ECs enhances survival, proliferation, migration, and differentiation of iNSPCs (Nakagomi et al., 2009a), indicating that augmentation of ECs (e.g., proliferation of ECs [angiogenesis]) can promote neurogenesis by enhancing the proliferation of endogenous iNSPCs.

Thus, therapeutic angiogenesis may enhance endogenous neurogenesis even after cerebral injury (Hamano et al., 2000; Chen et al., 2003). It has been reported that bone marrow cells (BMCs) such as bone marrow mononuclear cells (BMMCs) (Li et al., 2006; Kim et al., 2009a; Ribeiro-Resende et al., 2009) and mesenchymal stem cells (MSCs) (Labouyrie et al., 1999; Mahmood et al., 2004; Kurozumi et al., 2005) induce angiogenic effects by secreting multiple
growth factors including vascular endothelial growth factor (VEGF), glia-derived neurotrophic factor (GDNF), brain-derived neurotrophic factor (BDNF), nerve growth factor (NGF), and hepatocyte growth factor (HGF). We showed that BM-MCs can contribute to the proliferation of endogenous iNSPCs through vascular niche regulation, which includes EC proliferation following cortical infarction (Nakano-Doi et al., 2010).

In addition to ECs, astrocytes are also reported to be important niche cells for NSPCs in the SVZ (Song et al., 2002), SGZ (Lim et al., 1999) and cortex (Jiao et al., 2008). Our study already showed that astrocytes, as well as ECs, promote the proliferation of iNSPCs (Nakagomi et al., 2009a), suggesting that astrocytes function as a niche for cortex-derived iNSPCs. Although astrocytes were not observed within the post-stroke cortex after permanent ischemia (Nakagomi et al., 2011), astrocytes are resistant to hypoxia/ischemia and they can still survive after transient ischemia (Li et al., 1995). These findings might explain the reason why new-born neurons are frequently found in the post-stroke cortex after mild transient ischemia (Ohira et al., 2010), but are not seen after severe permanent ischemia (Nakagomi et al., 2009b).

Regulation of the immune system has also been proposed as one of the key factors in enhancing neurogenesis and functional recovery after stroke. Our studies showed that T lymphocytes, mainly CD4- but not CD8-positive T cells, induce apoptosis in iNSPCs (Saino et al., 2010; Takata et al., 2011). The details of the mechanism are still under investigation, but these findings suggest that the immune response and/or enhanced inflammation triggered by CD4-positive T cells, are major deteriorating modulators of post-stroke neurogenesis. These findings, at least in part, are consistent with previous results demonstrating that transplantation of mesenchymal cells accelerates endogenous neurogenesis after stroke (Li et al., 2008; Yoo et al., 2008), because such treatment is known to suppress the immune response in graft-versus-host disease.

2.3 Exogenous iNSPC transplantation after cerebral infarction

Compared to the strategy focusing on enhanced endogenous neurogenesis, exogenous NSPC transplantation may have some advantages in treating stroke patients; this therapy allows a longer therapeutic time window to administer larger numbers of stem cells, and to repeat the treatment. The therapeutic time window to enhance the endogenous neurogenesis may be limited, because we observed that neurogenesis peaks for several days and ends within a few weeks after stroke onset in patients (Nakayama et al., 2010).

Until now, various cell sources for exogenous NSPC transplantation have been proposed; e.g., fetal brain (Ishibashi et al., 2004; Kelly et al., 2004; Cayre et al., 2006; Darsalia et al., 2007), adult brain tissue obtained from the SVZ (Cayre et al., 2006; Hicks et al., 2007; Kameda et al., 2007), gene transfected bone marrow cells (Dezawa et al., 2004), immortalized tumor cell lines (Staines et al., 1994), embryonic stem (ES) cells/induced pluripotent stem cell (iPS) cells (Bjorklund et al., 2002; Wei et al., 2005; Buhmann et al., 2006) and ex vivo expanded cortex-derived iNSPCs (Nakagomi et al., 2009a; Nakagomi et al., 2009b; Nakano-Doi et al., 2010). Transplantation of exogenous NSPCs can be performed even at the chronic stage of post-stroke. In experimental models of stroke using fetal NSPCs, transplanted cells were reported to survive within the host brain, migrate into the injured area, and maintain their multipotency (Ishibashi et al., 2004; Kelly et al., 2004; Darsalia et al., 2007). However, there are
some issues to be solved for clinical application of exogenous NSPC transplantation in stroke patients; e.g., survival, safety and suitability of transplanted cells, and their capacity to repair injured adult brain. Indeed, the other lines of experiment using NSPCs derived from adult mammalian brains showed that only a small population of grafted cells can survive in the injured brain (Toda et al., 2001; Hicks et al., 2007; Kameda et al., 2007; Takahashi et al., 2008). Consistent with these reports, we showed that the majority of transplanted iNSPCs, which are derived from the adult cortex, cannot survive in the injured cortex (Nakagomi et al., 2009a).

A higher survival rate of transplanted NSPCs carrying the property of neoplasm (such as ES/iPS-derived NSPCs) can be expected, because survival is often attributed to a lack of apoptotic signaling. However, this property may be directly linked to a high risk of tumorigenesis. Whether the transplanted fetal NSPCs will be able to contribute to reconstitution of the adult brain is also an issue to be addressed, because they are the cells destined to form the infant brain. Therefore, we must achieve significant recovery of impaired neurological functions of the adult brain to determine the suitability of transplanted cells.

Our study showed that iNSPCs from the injured cortex differentiate into functional neurons with less tumorigenesis, suggesting that these cells are one of the most suitable NSPCs for transplantation. Therefore, we may choose alternative ways to continue the survival of transplanted iNSPCs. Recently, we reported that co-transplantation of iNSPCs with ECs as a vascular niche, enhances functional recovery after cortical infarction with longer survival of transplanted cells (Nakagomi et al., 2009a). This suggests that the microenvironment around the transplants has to be considered for cell therapy. From another point of view, as differentiated cells are more resistant to apoptotic cell death, enhancing differentiation of NSPCs into mature neurons may be a choice in maintaining the transplant. Recent studies showed that transplantation of NSPCs with valproic acid, which inhibits proliferation but enhances differentiation of transplanted stem cells to functional neurons, significantly improves motor function in a spinal cord injury model (Abematsu et al., 2010). These results may indicate a future direction for the clinical application of exogenous NSPC transplantation for patients after cerebral infarction.

Another problem regarding cell transplantation is the difficulty in regulating the differentiation of transplanted NSPCs in vivo. It is well-known that a variety of chemical mediators/cytokines are produced/activated at the site of brain injury, and among these, IL-6, CNTF, and BMPs promote differentiation of NSPCs into the astrocytic phenotype (Nakashima et al., 1999; Okada et al., 2004). Our previous studies showed that transplanted iNSPCs largely differentiated into glial cells in vivo, although they predominantly differentiated into neuronal cells in vitro (Nakagomi et al., 2009a). These results suggest that the neurogenesis-oriented regulation of transplanted iNSPCs might accomplish a real functional restoration of stroke patients in the future.

3. Conclusion

In conclusion, we demonstrated that iNSPCs, which are the potential cell sources for neocortical neurogenesis, develop in the murine post-stroke cortex (Nakagomi et al., 2009a; Nakagomi et al., 2009b; Nakano-Doi et al., 2010; Saino et al., 2010). Furthermore, we
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This book is a collective work of international experts in the neural stem cell field. The book incorporates the characterization of embryonic and adult neural stem cells in both invertebrates and vertebrates. It highlights the history and the most advanced discoveries in neural stem cells, and summarizes the mechanisms of neural stem cell development. In particular, this book provides strategies and discusses the challenges of utilizing neural stem cells for therapy of neurological disorders and brain and spinal cord injuries. It is suitable for general readers, students, doctors and researchers who are interested in understanding the principles of and new discoveries in neural stem cells and therapy.

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