LNCaP Prostate Cancer Growth In Vivo: Oncostatic Effects of Melatonin as Compared to Hypoxia and Reoxygenation

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1. Introduction

Oxygen (O\textsubscript{2}) is often thought of as a double-edged sword: all forms of life actually need O\textsubscript{2} for survival as the terminal acceptor of electrons in oxidative phosphorylation, but excess O\textsubscript{2} might increase formation of reactive O\textsubscript{2} species (ROS). Whereas on one hand ROS trigger uncontrolled burst of free radicals that lead to potentially lethal injury, on the other hand they act as messengers that elicit cell protection and improve survival through a variety of mechanisms. Although one can easily expect a link of O\textsubscript{2} with tumor growth and metastatic potential, there is no univocal role for O\textsubscript{2} in cancer.

1.1 Hypoxia in cancer

Lack of O\textsubscript{2}, or hypoxia, has been extensively studied in cancer because the growth of solid tumors requires a local vascular network that supplies O\textsubscript{2} and nutrients to tumor cells. In the classical view, when cell proliferation exceeds angiogenesis, the vasculature might become unable to sustain the O\textsubscript{2} needs of tumor cells, which therefore have to cope with an environment chronically deficient in O\textsubscript{2} as a result of diffusion-limited O\textsubscript{2} supply (Vaupel, 2004). This triggers various mechanisms, most of which are mediated by over-expression of the hypoxia-inducible factor-1 (HIF-1\textalpha). HIF-1\textalpha stimulates a variety of mechanisms aimed at survival of hypoxic tissue, for example the angiogenic switch, which provides growth factors for the development of circulation to feed the growing tumor, as well as anti-apoptotic and cell cycle factors. Therefore, tumor hypoxia emerges as a major contributor to the malignant phenotype (Hockel et al., 2001) and a cause of resistance to radiation therapy (Brown et al., 1998).

There are several instances whereby the O\textsubscript{2} supply to tissues is altered. The term “hypoxia” is sometimes improperly attributed to any of them, but there are important differences with dramatically different phenotypes, as explained in Table 1.

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Prostate cancer is the most common neoplasia and the second most frequent cause of male cancer death in the developed world and in many Western countries (Hsing et al., 2000). A malignant tumor derived from the interaction of genetic and environmental factors, potentially curative treatment options are available for management of early localized disease, whereas palliative hormonal therapy in the form of medical or surgical castration is the mainstay of treatment for patients with advanced prostate cancer. Approximately 80% of castrated patients will suffer from a relapse of the disease within 2 years, with progression of the tumor from a hormone-dependent to a hormone-independent stage (Wilding, 1995), which is associated with unfavorable prognosis.

In addition, prostate cancer may also represent an useful workbench to investigate the relationship between hypoxia and cancer, because HIF-1α is over-expressed compared with normal prostate epithelium (Zhong et al., 1999) and its up-regulation is recognized as an early event in carcinogenesis (Zhong et al., 2004). Furthermore, androgens and androgen receptors modulate HIF-1α levels (Kimbro et al., 2006), and hypoxia increases androgen receptor activity in LNCaP cells in vitro via HIF-1α (Park et al., 2006).

1.3 Melatonin and prostate cancer

Melatonin (N-acetyl-5-methoxytryptamine, Figure 1), a neurohormone synthesized during night time in the pineal gland, mediates many physiological, endocrinological and behavioral processes, including the well-known biorhythmic regulation of organism physiology, through its action on the biological clock at the hypothalamic suprachiasmatic nucleus (Tamarkin et al., 1985). Melatonin increases sleepiness, decreases core temperature, and increases peripheral temperature in humans (Brzezinski, 1997; Burgess et al., 2001; Lewy et al., 1996). The melatonin’s regulatory roles are mediated through high affinity G protein-coupled receptors that reside primarily in the eye, kidney, gastrointestinal tract, blood...
vessels, and brain (Beyer et al., 1998). This suggests some significant actions of melatonin on the cell biology of these target tissues outside the central nervous system as well (Cardinali et al., 1997; Pang et al., 1993). In addition, melatonin is known to exhibit antioxidant properties against the deleterious effects of reactive oxygen and nitrogen species (ROS and RNS, respectively) that are independent of its many receptor-mediated effects (Korkmaz et al., 2009; Ochoa et al., 2011). Melatonin has been reported to scavenge hydrogen peroxide (H₂O₂), hydroxyl radical (HO•), nitric oxide (NO•), peroxynitrite anion (ONOO⁻), hypohlorous acid (HOCl), singlet molecular oxygen [O₂(¹Δg)] and superoxide anion (O₂⁻) (Allegra et al., 2003; Reiter, 1998; Reiter et al., 2001; Tan et al., 2000). Melatonin has also been shown to possess genomic actions, through regulation of the expression of several genes including glutathione peroxidase, superoxide dismutase, and catalase, both under physiological conditions and under conditions of elevated oxidative stress (Allegra et al., 2003; Kotler et al., 1998).

Fig. 1. Structure of melatonin, MW 232.2 Da.

A multitude of literature reports have documented a direct modulatory effect of melatonin on benign and malignant cell proliferation and an anti-tumor effect was reported both in vitro and in vivo in human tumors of the reproductive tissues (Cos et al., 1998; Shiu et al., 1999). Among these, a significant role of melatonin, by itself and by interaction with sex steroids, has been found in the pathobiology of prostate cancer and benign prostatic hyperplasia (Laudon et al., 1996; Lupowitz et al., 1999). Of the various experimental prostate cancer models, hormone (androgen)-sensitive and hormone (androgen)-insensitive metastatic human prostate cancer cell lines are widely used for the experimental evaluation of pharmacological agents with therapeutic potential for the disease (Cho et al., 2011). In particular, the growth of the androgen-independent but androgen-sensitive (responsive) human LNCaP prostate cancer cells has been demonstrated to be inhibited by the pineal gland indoleamine hormone both in vitro and in vivo in a nude mice xenograft model (Siu et al., 2002; Xi et al., 2000). The antiproliferative action of melatonin seems to be mediated in part by means of MT1 receptor activation and partly by means of attenuation of dihydrotestosterone-induced calcium influx model (Xi et al., 2000).

1.4 Solid lipid nanoparticles and cryopass laser therapy
Cytostatic and antitumoral drugs activity is often impaired by low plasma solubility, poor systemic absorbance, rapid metabolism, non-specific tissue distribution and toxic effects. The search for advanced methods to deliver these molecules to target tissues and to overcome the failure shown even with very active drugs is therefore mandatory.
Solid-lipid nanoparticles (SLN) is a technology able to produce sub-micrometric lipidic particles characterized by an average diameter <500 nm, with a narrow size distribution and a spherical shape. The several advantages of this technology are listed in Table 2 (Mehnert et al., 2001). SLN formulation of antitumor drugs is predicted to be advantageous with respect to other formulations because it allows greater uptake into the malignant cell, with consequent intracellular accumulation and higher efficacy, still keeping the level of free circulating drug as low as possible, thereby preventing non-specific unwanted side effects.

<table>
<thead>
<tr>
<th>Main advantages of SLN formulation for drugs delivery.</th>
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<tbody>
<tr>
<td><strong>To control and direct the release of the drug against a specific target organ</strong></td>
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<tr>
<td><strong>To modify favorably drugs pharmacokinetics</strong></td>
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<tr>
<td><strong>To target directly to lymph bypassing the entero-hepatic circulation following oral administration</strong></td>
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<tr>
<td><strong>To obtain a stealth formulation that avoids recognition by the reticuloendothelial system</strong></td>
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<tr>
<td><strong>To allow oral administration of drugs poorly soluble and not easily adsorbed in the gastrointestinal tract</strong></td>
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<tr>
<td><strong>To overcome the blood-brain barrier</strong></td>
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<tr>
<td><strong>To permit administration routes as transdermal or eye topical</strong></td>
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Table 2. Main advantages to administer drugs as solid-lipid nanoparticles (SLN).

Criopass therapy, is a procedure used to actively deliver drugs across the dermal barrier. It is based on topical application of a frozen drug emulsion in 1.5% hydroxymethylcellulose by means of a laser source that gives energy to penetrate the dermal barrier and deliver the active principle to the target area. The low energy photon flux generated by a laser beam hit the drug molecules frozen in the crystal lattice, exciting the electrons in the outer orbital. The drug molecules, when melt at the ice-skin interface, release the accumulated potential energy transforming into kinetic energy, which speeds up the passage of the drug across the skin membrane and allows to reach the target area. The last step consists in a laser scan on the area of drug application to optimize the adsorption through the skin and facilitate the drug to reach the desired site of activity. This treatment is particularly advantageous for treatment of bones cartilage, producing significant drug accumulation in a tissue difficult to be reached by traditional administration techniques. The main advantages of this non-invasive and painless treatment are speed of absorption (15-20 s), high capacity of penetration (6 cm depth), suitability for polar and non-polar molecules delivery, improved drug bioavailability and high specificity.

1.5 Aims

Solid tumors contain underperfused regions where hypoxia might induce adaptation and cell proliferation. Often, this response is mediated by HIF-1α over-expression in hypoxic
cells. We demonstrated that systemic chronic in vivo hypoxia promotes prostate cancer growth regardless of HIF-1α expression level and neovascularization. We have also assessed that altering HIF-1α expression by use of non-pharmacological agents (intermittent hypoxia with reoxygenation) alter the phenotype of tumor growth. These observations suggest an important role for hypoxia dependent pathways that do not involve HIF-1α, and for the pharmacological treatments able to modulate these pathways.

Pharmacologic concentrations of melatonin inhibit in vitro expression of HIF-1α protein under both normoxic and hypoxic conditions in DU145, PC-3, and LNCaP prostate cancer cells (Park et al., 2009). This effect, perhaps a result of the antioxidant activity of melatonin against ROS induced by hypoxia, and the subsequent suppression of HIF-1α transcriptional activity decreases VEGF expression in HCT116 human colon cancer cell line (Park et al., 2010). Therefore, the main aim of this preliminary study was to focus into the role of melatonin as a therapeutic/adjuvant agent that reduces tumor growth in vivo as a proof-of-concept for further studies assessing melatonin interference with the hypoxia signaling paths. To this purpose, we used the in-vivo model of nude mice xenograft with human LNCaP prostate cancer cells under a variety of conditions spanning from chronic hypoxia with/out reoxygenation, and compared different routes of melatonin administration.

2. Methods

Cells. LNCaP cells (80-90% confluence) were maintained in RPMI-1640 medium containing 10% (v/v) heat-inactivated fetal bovine serum and L-glutamine, and cultured in 5% CO2. To ensure that LNCaP cells were not injured when passing through G26 needles during xenografts, we verified that their vitality did not decreased by more 2-3% per each passage. To obtain positive controls for HIF-1α immunostaining, LNCaP cells were incubated for 24 h in the presence of 100 μM CoCl2, washed, fixed in formalin and stained as described below.

Mice. Seven-week old Foxn1nu/nu mice (Harlan, n=46), weighing 25-30 g at the entry into the study, were cared in accordance to the Guide for the Care and Use of Laboratory Animals published by the National Institutes of Health (NIH Publication No. 85-23, revised 1996). Water and bedding were heat-sterilized, whereas food was sterilized by 60Co γ-irradiation. Mice had free access to water and conventional laboratory diet until 24 h before sacrifice. A 12/12 h light/dark cycle was maintained.

Xenografts. LNCaP cells were resuspended in ice-cold Matrigel (1:1) at a final concentration of 3×10⁶/0.1 ml. Mice were inoculated in each flank with 0.1 ml of cells using a 26G insulin syringe. The next day, mice were transferred into the gas chamber, where they were treated accordingly.

In vivo measurements of tumor growth. Body weight and tumor volume were measured three times a week during the various treatments. The tumor volume was calculated as length•width•height•0.5236, as measured by a caliper. Data are expressed as the ratio (tumor volume)/(body weight) to compensate different rates of growth in the various experimental situations (Figure 2).

Sacrifice. At the end of the observation period, mice were anesthetized by i.p. Na-thiopental (10 mg/100 g body weight) plus heparin (500 units), then they were thoracotomized to
withdraw a blood sample into a heparinized syringe from the left ventricle, and tumors were quickly excised from surrounding skin.

Fig. 2. Picture of a mouse at the end of the observation window. The tumors are clearly visible on both flanks.

*Hemoglobin.* Hemoglobin (Hb) concentration was measured in blood, by diluting 10 µl of well-stirred blood in 1 ml of Drabkin reagent, followed by incubation for 30 min at room temperature and absorbance reading at $\lambda=540$ nm. The concentration was calculated assuming $\lambda=11.05$ cm$^{-1}$ mM$^{-1}$.

*Effects of melatonin i.p.* For this set of experiments, mice xenografted with LNCaP cells as described above were treated with melatonin i.p. (30 µg, 1 mg/Kg), given either dissolved in in 100 µL isotonic saline (n=7) or encapsulated in 100 µL SLN (n=7). Mice treated with 100 µL saline acted as control. Whereas melatonin-saline was prepared fresh, melatonin-SLN was obtained from Nanovector S.r.L., Torino, Italy (Gasco et al., 2007). Timing of treatments is given in *Figure 3*.

Fig. 3. Timing of melatonin treatments.

*Effects of melatonin-laser.* For this set of experiments, mice xenografted with LNCaP cells as described above were exposed to laser treatment with (n=11) or without (n=11) melatonin. Melatonin was prepared fresh every week by emulsifying 0.048 mg melatonin/mL of 1.5 % hydroxymethylcellulose for 7 min with a Ultraturrex at the maximum speed in ice and dark.
Then, 15 mL of the suspension or 0.72 mg melatonin was transferred in suitable devices and frozen at -20°C overnight. Each frozen stick (Figure 4A) was used to treat 6 mice. The final dose administered topically was about ~0.120 mg melatonin/mouse/treatment, i.e. 4 mg/Kg. For the administration, the stick containing frozen melatonin (or saline for the control group) was delivered topically for 2.4 min in the correspondence of the xenografts with the frozen stick connected to the laser beam. The duration of the treatment was kept within 2.4 min, because topical application of the frozen stick in small-size animals causes hypothermia (when necessary, mice were kept on a heating plate at 37°C during the treatment, not shown). The concentration of melatonin was selected in order to deliver the wanted amount in 2.4 min. After this first phase, mice were placed in a home-made device, immobilized and exposed for 15 min to the laser scan (the wide of the laser beam scan was set to the minimum value, so to cover only the small tumor area exposed on the mice back) (Figure 4B). We used an instrument LASERICE Med C.I.R.C.E. S.r.L., Magnago, Milano (230 V, 50 Hz, 150 mA) constituted by a device for freezing the drug emulsion and by a scanner connected to photodiode laser bean with $\lambda = 635$ nm, maximum power <5 mW, collimation lens <20 mV. For our experiments, we selected the software designed for veterinary use in small animals. The software automatically selects the most suitable laser power and frequency to target the drug at the right depth into the tissues.

Fig. 4. (A) Topical administration of melatonin by a frozen stick connected to a low energy laser beam. (B) Experimental setup showing exposure of immobilized mice to high-energy laser scan.

**Effects of hypoxia with/out reoxygenation.** Data relative to this set of experiments have been published in (Terraneo et al., 2010) and are here reported to enable comparing with the groups with melatonin. Briefly, mice xenografted with LNCaP cells as described above were exposed to either chronic hypoxia (10% O$_2$) or hypoxia with reoxygenation (3 times/week for 1 h), with normoxia as control (n=17, 19 and 20, respectively). Hypoxia was induced by using the hypoxic chambers described elsewhere that prevent any unwanted contact of the animal with room air during cleaning operations and sacrifice (Milano et al., 2002).
Statistics. Data are expressed as mean ± SEM. Significance level was P=0.05 (two-tailed). To detect differences among the groups, we performed one-way ANOVA. If this test resulted significant, the differences between selected pairs of data were tested using the Bonferroni procedure (Instat 3, GraphPad software).

3. Results

Table 3 shows the main characteristics of the mice considered in this study. No significant differences were detected among the various melatonin groups as far as the changes in body weight and Hb concentration are concerned. By contrast, hypoxia depressed the gain in body weight and increased the blood Hb concentration, as expected. The marked decrease in body weight in the mice exposed to hypoxia forced us to shorten the 42-day observation window to 28 days.

<table>
<thead>
<tr>
<th></th>
<th>n Surviving/total</th>
<th>Body weight at entry, g</th>
<th>Final body weight, g (42 days of treatment)</th>
<th>Blood [Hb], g/L</th>
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<tr>
<td><strong>Effects of melatonin i.p.</strong></td>
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<tr>
<td>Saline</td>
<td>6/6</td>
<td>26.63±0.20</td>
<td>26.77±0.80</td>
<td>120±3</td>
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<tr>
<td>Melatonin-saline</td>
<td>7/7</td>
<td>27.71±0.32</td>
<td>32.17±0.46*#</td>
<td>119±6</td>
</tr>
<tr>
<td>Melatonin-SLN</td>
<td>7/7</td>
<td>27.96±0.25</td>
<td>31.94±0.90*#</td>
<td>119±3</td>
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<tr>
<td><strong>Effects of melatonin laser</strong></td>
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<tr>
<td>Laser</td>
<td>11/11</td>
<td>27.24±0.31</td>
<td>29.70±0.91*</td>
<td>126±11</td>
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<tr>
<td>Laser+melatonin</td>
<td>11/11</td>
<td>27.40±0.30</td>
<td>30.58±0.74*</td>
<td>119±6</td>
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<tr>
<td><strong>Effects of hypoxia (28 days of treatment)</strong></td>
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<td></td>
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<tr>
<td>Normoxia</td>
<td>19/20</td>
<td>26.83±0.39</td>
<td>31.17±0.61*</td>
<td>134±3</td>
</tr>
<tr>
<td>Chronic hypoxia</td>
<td>16/17</td>
<td>28±0.46</td>
<td>27.38±0.52#</td>
<td>197±3#</td>
</tr>
<tr>
<td>Hypoxia with reoxygenation</td>
<td>9/9</td>
<td>27.55±0.37</td>
<td>23.92±0.28*#</td>
<td>205±5#</td>
</tr>
</tbody>
</table>

Table 3. Main characteristics of the mice considered in this study. Data from the hypoxia groups are published in (Terraneo et al., 2010). *, P<0.05 with respect to body weight at entry (unpaired two-tailed Student’s t-test); #, P<0.05 with respect to the relative control (ANOVA and Bonferroni post-test).

3.1 Effect of melatonin i.p.

This set of experiments aims at assessing whether the traditional way to administer melatonin results into an oncostatic situation, and whether administration of the same amount of melatonin is oncostatic as well. The xenograft rate of success was in the range 55-90%. Whereas Figure 5A reports the time course of the successful xenografts, Figure 5B reports the time after the xenograft necessary for the xenografted tumors to be palpable. Data confirm that i.p. treatment with melatonin in saline has important oncostatic potential. Of interest, melatonin does not delay appreciably the time of appearance of the tumors after the xenograft, but rather it decreases the growth rate. Melatonin in SLN has less oncostatic potential than melatonin in saline when given in the same amount, probably as the result of less delivery efficiency than in the melatonin-saline group.
Fig. 5. Time course (A) and time necessary for the tumors to be palpable (B) in mice treated i.p. with either melatonin in saline or melatonin in SLN (0.51 mg melatonin/mouse in 17 administrations over a 42 day period), whereas saline-treated mice represent the control group. Data are expressed as mean ±SEM from 12, 10 and 8 tumors, respectively. *, P<0.05 from untreated; #, P<0.05 from melatonin SLN (ANOVA and Bonferroni post-test).

3.2 Effect of melatonin laser
This set of experiments aims at assessing whether administering melatonin topically by treatment with laser results into an oncstatic situation as that described above for i.p. melatonin. The xenograft rate of success was in the range 75-95%. Whereas Figure 6A reports the time course of the successful xenografts, Figure 6B reports the time after the xenograft necessary for the xenografted tumors to be palpable. Whereas laser+melatonin results into a oncostatic situation (compare with the saline-treated as reported in Figure 4), it is difficult to discern an effect due to melatonin in comparison with that driven by laser+melatonin.

Fig. 6. Time course (A) and time necessary for the xenografted tumors to be palpable (B) in mice exposed to either laser only or laser+melatonin (2.04 mg melatonin/mouse in 17 administrations over a 42-day period). Data are expressed as mean±SEM from 12, 13 and 12 tumors, respectively. No significant difference between the two treatments was observed, but both laser groups are different from Saline control (*, P<0.05).
3.3 Effect of hypoxia
This set of experiments aims at assessing whether inducing local hypoxia increases the growth of prostate tumors as published elsewhere (Terraneo et al., 2010). By decreasing arterial PO$_2$ from 85 to 34 mmHg, the selected hypoxia severity (10% O$_2$, equivalent to an altitude of about 5000 m) decreases the total O$_2$ arterial content by 35%, despite the increased blood Hb concentration. The xenograft success rate was 56-83% without any significant effect of hypoxia nor its way of administration. Figure 7A reports the time course of the successful xenografts, and Figure 7B reports the time after the xenograft necessary for the tumors to be palpable. Chronic hypoxia resulted into significant increase of the tumor growth rate. Interestingly, an operation aimed at increasing the cytosolic abundance of HIF-1α by approximately 10 times, e.g., exposing mice to repeated reoxygenation events during hypoxia, did not affected appreciably the tumor growth rate. This suggests that mechanisms other than those mediated by HIF-1α might have determined the increased tumor growth rate in hypoxic mice.

![Fig. 7. Time course (A) and time necessary for the xenografted tumors to be palpable (B) in mice exposed to either laser only or laser+melatonin (n g melatonin/mouse in n administrations over a 42 day period). Data are expressed as mean±SEM from n, n and n tumors, respectively. *, P<0.05 from normoxia (ANOVA and Bonferroni post-test).](image)

4. Discussion
Melatonin. After the first observation that in MCF-7 cells in vitro, 1 nM melatonin reduces tumor cells invasiveness in Falcon invasion chambers (Cos et al., 1998), the oncostatic properties of melatonin have been thoroughly investigated. Melatonin acts synergistically with castration in inhibiting growth of androgen-sensitive LNCaP tumor through opposite changes in cyclin D1 levels induced by activated MT1 and EGF receptors (Siu et al., 2002). As MT1 receptors are clearly involved in androgen-sensitive LNCaP, but not in androgen-insensitive PC-3 cells (Xi et al., 2001), it is likely that the antiproliferative action of melatonin in LNCaP tumor growth is associated with MT1 receptor protein expression. Furthermore, the clear relationship found in MCF-7 xenografts between melatonin and telomerase activity, responsible of telomere elongation that activated in most human cancers (Leon-Blanco et al., 2003), suggests that melatonin also influences telomerase decreasing its activity in the tumors. Although melatonin at physiologic concentration has no impact on VEGF
expression in three human cancer cell lines (PANC-1, HeLa and A549), at high pharmacological concentrations it markedly reduces expression of VEGF and HIF-1α induced by CoCl₂ in cultured cancer cells (Dai et al., 2008). A breakthrough in understanding the mechanisms underlying the oncostatic properties of melatonin came along with unraveling the relationship between melatonin and the hypoxia signaling path. Melatonin indeed inhibits HIF-1α protein expression in normoxic and hypoxic DU145, PC-3 and LNCaP prostate cancer cells without affecting HIF-1α mRNA levels (Park et al., 2009). In HCT116 human colon cancer cells, melatonin destabilizes HIF-1α, suppresses its transcriptional activity, thereby decreasing VEGF expression (Park et al., 2010). In turn, these features block in vitro tube formation and invasion and migration of human umbilical vein endothelial cells induced by hypoxia media, indicating that melatonin plays a pivotal role in tumor suppression via inhibition of HIF-1α-mediated angiogenesis. Our observation that melatonin inhibits LNCaP prostate cancer growth as opposite to enhanced growth in hypoxia agrees with the described findings.

Chronic hypoxia. Despite the acknowledged role of hypoxia in cancer biology, mainly acquired through in vitro and clinical studies, accurate analysis of existing literature revealed that the effects of chronic hypoxia in vivo had not been investigated experimentally as well. To mimic chronic hypoxia, we recently developed an experimental model whereby prostate tumor-bearing mice are exposed to various forms of hypoxia in vivo, while continuously monitoring tumor growth and finally assessing the molecular and cellular phenotypes (Terraneo et al., 2010). We found that, although hypoxia in vivo promotes prostate cancer growth, this could not be entirely ascribed to HIF-1α, in the favor of other molecular pathways, such as those involving phosphatidylinositol-3-phosphate/protein kinase B (Akt) pathway. This finding bears important implications especially when designing effective therapies: as a matter of facts, on clinical ground therapies targeting HIF-1α do not appear particularly successful (Fox et al., 2011). Comparing the effects led by chronic hypoxia with/out reoxygenation contributed to univocally assess the latter issue.

Chronic hypoxia with reoxygenation. Reoxygenating the tumors during hypoxia increased HIF-1α cytosolic level >10-fold more than in chronic hypoxia, yet tumor growth was essentially similar to that of chronic hypoxia (Terraneo et al., 2010). Higher HIF-1α in reoxygenated tumors with respect to chronic hypoxia could be a consequence of shorter normalization time since the last exposure to hypoxia (2-3 days), or of reoxygenation-induced HIF-1α stabilization or enhanced synthesis. As the reoxygenation of hypoxic tissues causes considerable oxidative stress (Milano et al., 2004), the associated enhanced generation of mitochondrial ROS may stabilize HIF-1α (Chandel et al., 2000). It has also been proposed that the persistent oxidative stress promoted by ROS during the reoxygenation further amplifies HIF-1α activation in a feed-forward mechanism through a mechanism involving mTOR (Semenza et al., 2007). Therefore, although this finding was key to suppose non-centrality of HIF-1α in cancer growth, the effect of HIF-1α on metastatic potential is still to be investigated.

Therapeutic potential and future perspectives. The role of melatonin as anti-proliferative factor in the management of prostate cancer is currently under study by several research groups. However, its signaling mechanism and its crosstalk with other positive or negative growth regulator factors as HIF-1α, sex steroids, epidermal growth factor (EGF), vascular endothelial growth factor (VEGF) and others is still under debate and deserves to be thoroughly investigated. Moreover, to elucidate the biological effects of melatonin under
physiological and pathological conditions, its putative antioxidant action is to be studied. Finally, the effect of melatonin on sarcosine (N-methyl derivative of the amino acid glycine), a potentially important metabolic intermediary of cancer cell invasion and aggressivity (Sreekumar et al., 2009) merits future attention.

5. Conclusion

In vivo systemic hypoxia promotes prostate cancer growth regardless of HIF-1α expression level and neovascularization, suggesting an important role for hypoxia-dependent pathways that do not involve HIF-1α, as the phosphatidyl inositol-3-phosphate signaling cascade. Melatonin experiments may not only provide a useful probe to assess the effects of HIF-1α on tumor growth, but may also represent the basis for the future introduction of this natural molecule as adjuvant active component in novel therapeutic strategies for the treatment of malignant prostate cancer in humans, for the prevention of cancer relapses, or simply for the amelioration of the quality of life of the oncologic patient.

6. Acknowledgments

We thanks to Dr. Enrico Bonizzoni and Dr. Emilio Bonizzoni C.I.R.CE. S.r.L., Magnago, Milano, who kindly borrowed the Laser equipment.

7. References


This book encompasses three sections pertaining to the topics of cancer biology, diagnostic markers, and therapeutic novelties. It represents an essential resource for healthcare professionals and scientists dedicated to the field of prostate cancer research. This book is a celebration of the significant advances made within this field over the past decade, with the hopes that this is the stepping stone for the eradication of this potentially debilitating and/or fatal malignancy.

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