

Spices as Alternative Agents for Gastric Ulcer Prevention and Treatment

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1. Introduction

1.1 Important aspects on ulcer pathogenesis

World-wide, peptic Ulcer disease (PUD) is considered as a common gastrointestinal disorder. It develops as a result of altered balance between offensive and defensive factors. Offensive (aggressive) factors disrupt normal mucosal integrity and allow H⁺ back diffusion with a subsequent cellular injury. *Helicobacter pylori* (*H. pylori*) and nonsteroidal anti-inflammatory drugs (NSAID) represent the major aggressive factors associated with PUD. Experimentally induced gastric ulcer has expanded our knowledge on ulcer pathogenesis. Indomethacine, 80% ethanol and pyloric ligation are the methods commonly applied in experimental ulcer models. Other universally accepted experimental ulcer models include 0.2 mol/L NaOH, 25% NaCl, stress induced by swimming (1), acetylsalicylic acid (2), cold-restraint (3) and hypothermic restraint (4).

A major event in the pathogenesis of NSAID induced gastric ulcer is represented by inhibition of prostaglandin (PG) synthesis, enhancement of gastric acid secretion, suppression of bicarbonate secretion, glutathione (GSH) levels, mucosal circulation, cell proliferation and growth as well as alteration of gastric mucosal barrier integrity. Inhibition of PG biosynthesis enhances generation of leukotrienes and other products of the 5-lipoxygenase pathway (5). These products disrupt the mucosal barrier with subsequent enhancement of gastric mucosal permeability for H⁺ ions and Na⁺ ions and reduction of transmucosal potential difference (6, 7). Furthermore, NSAID uncouple mitochondrial oxidative phosphorylation, affect mitochondrial morphology, reduce the intracellular ATP levels and alter the normal regulatory cellular function (8). These processes promote erosions and ulcer formation. In addition, generation of reactive oxygen species (ROS) is also considered as a major factor contributing to ulcer pathogenesis. Another, prostaglandin-independent pathway of gastric ulcer pathogenesis is induced by enhanced endothelial adhesion, activation of polymorphonuclear cells (PMN) with subsequent release of oxidative byproducts (9, 10). PMN activation induces depletion of GSH and sulfhydryl compounds (SH) in tissue with enhanced mucosal myeloperoxidase (MPO) and malondialdehyde (MDA) concentration (11). Myeloperoxidase is considered as a marker of oxidative process induced by PMN tissue infiltration.

Similarly, in ethanol-induced gastric mucosal injury there is enough evidence to suggest the role of oxidative burst. Ethanol-induced oxidative damage is commonly associated with

generation of ROS, leading to oxidative stress. Acute ethanol treatment induced oxidative damage is associated with a decreased GSH content in gastric tissue along with an increased MDA and xanthine oxidase activity (12). The oxygen free radicals-induced lipid peroxidation affects mitochondrial energy metabolism and plays a critical role in the pathogenesis of acute ethanol-induced gastric mucosal injuries (13). On the mitochondrial level, ethanol-induced intracellular oxidative stress causes also mitochondrial permeability transition with mitochondrial depolarization that precedes gastric mucosal cells necrosis. This process can be prevented by intracellular antioxidants, such as GSH (14). Acute ethanol administration is also associated with inhibition of catalase and, glutathione peroxidase (GPx) activities with significant increase of MDA contents, MPO activity, and cellular apoptosis (15). Furthermore, ethanol induces inhibition of SH with enhancement of superoxide dismutase (SOD) and glutathione reductase (GR) activities (16). The extent of oxidative damage in stomach as indicated by the ulcer index, gastric mucosal MDA content and alteration of mitochondrial ultrastructure is correlated with ethanol exposure and concentration (13).

2. Introduction in spices

Spices are used in several parts in the world as food additives and carminatives. Since ancient times they are also applied in the traditional management of a variety of disorders. Currently their therapeutic value has gained a considerable interest and several investigators have reported their effects in laboratory animals and in man. It has been experimentally demonstrated that spices, herbs, and their extracts possess antibacterial (17) antifungal (18,19), vermifugal, nematocidal, molluscicidal properties (20-22), anti-inflammatory and antirheumatic activity (23-25), hepatoprotective (26,27), nephro-protective (28), antimutagenic, anticancer potentials (28-30) and antihypercholesterimc potentials (31-34).

A tremendous number of studies have evaluated the antiulcer effect of spices. Although some investigators have reported deleterious effect of certain spices such as red and black pepper, the majority have demonstrated rather a cytoprotective activity in animal (35-37) as well as in human (38).

3. Factors involved in ulcer healing

In the presence of mucosal barrier disruption, several factors including acid, bile acids, NSAID and ethanol promote H⁺ back diffusion and enhance the susceptibility to develop ulcer. On the other hand, optimal mucosal microcirculation and bicarbonate secretion with formation of an alkaline buffer layer at the epithelial surface is considered as a first line of mucosal defense and gastroprotection. Other factors including prostaglandins (PG), growth factors (GF), nitric oxide (NO) or calcitonin gene-related peptide (CGRP), as well as some gut hormones such as gastrin, cholecystokinin (CCK), leptin, ghrelin, gastrin-releasing peptide (GRP) and melatonin are involved in mucosal defense system and the ulcer healing process. The protective action of gut hormones is attributed to the release of cyclooxygenase-2 (COX-2) and PGE₂ at the ulcer margins (39, 40) or activation of sensory nerves (41). In addition, Tumor necrosis factor - α (TNF- α), released during gastric mucosal injury, activates PG pathway and promotes epithelial cell repair and healing (42). EGF and other growth factors are also pivotal for the process of mucosal healing. Furthermore, in response to gastric injury and inflammation, gastrin and parietal cells contribute to the regulation of mucosal proliferation (43).

4. Why spices, herbs and plant extracts are considered as an alternative ulcer therapy?

Ulcer healing and prevention of recurrence represent the central goals of treatment. The treatment is targeted at either counteracting aggressive factors like acid, pepsin, active oxidants, platelet aggravating factor (PAF), leukotrienes, endothelins, bile or exogenous factors including (NSAID) or enhancing mucosal defense such as mucus, bicarbonate, blood flow, PG and NO (44). Antisecretory drugs including H₂-receptor antagonists (H₂-RA) and proton pump inhibitors (PPI) alone or combined with antibiotics in the presence of *H. pylori* infection are currently considered as the most acceptable drugs for ulcer treatment. The main action of antisecretory drugs is acid suppression. These agents lack effect on other factors involved in ulcer pathogenesis and therefore, do not meet all treatment goals. In addition, acid suppressors are expensive and associated with adverse effects and ulcer recurrence. Hence, efforts are on to search for suitable alternative treatment from medicinal plants resources. Already a large percentage of world population relies on medicinal plants to treat a variety of disorders including PUD. In addition to their ability to act on various pathogenetic factors, they are cheap and easily accessible. Furthermore, a large number of spices and plant extracts evaluated by various researchers for their anti-ulcer effects have a favorable outcome. (29, 45-48)

The antiulcer effect of spices/herbs is based on the activities of their chemical constituents, which attenuate the gastric secretion, enhance mucosal integrity, interfere with oxidative burst, NO, SH compounds and inhibit *H. pylori* growth. Due to their variable phytochemical constituents they may exhibit antisecretory, cytoprotective, antioxidant or combined activities.

5. Herbs and gastric secretion

A variety of spices and their extracts possess a potent antisecretory activity. Pylorus-ligation in rats represents the model of antisecretory studies. A number of spices, herbs and plants methanolic or aqueous extracts possess an antisecretory activity. For instance, *Cissus quadrangularis*, *Maytenus ilicifolia*, phytosphingosine, *Cecropia glaziovii* Sneath (*Cecropiaceae*), alkaloid extract and 2-phenylquinoline obtained from the bark of *Galipea longiflora* (*Rutaceae*) and *Landolphia owariensis* induce significant inhibition of acidity, pepsin content and ulcer index (49-54), respectively. This potent antisecretory action of spices and plant extracts is likely related to their flavonoid content. *Maytenus ilicifolia* is considered among flavonoid-rich plant extracts (55).

In rats with pylorus ligation, antisecretory effect of methanolic extract of *Momordica charantia* L. is demonstrated by decrease in acidity, pepsin content and ulcer index with an increase in gastric mucosal content (56).

Likewise, the protective effect of *Cissus quadrangularis* extract is mediated by inhibition of gastric secretion with decrease in ulcer index as well as enhancement of mucosal defense (49).

6. Herbs and cytoprotection

Several spices and plants extracts promotes ulcer healing via enhancing gastric mucosal content beside their antisecretory activity in pylorus-ligated rats. Ginger rhizome extract-induced cytoprotective activity is based on its antioxidative and gastric mucosal protective

activities (1). Total carotenoid and astaxanthin esters protect mucin, enhance antioxidants enzymes level and H⁺, K⁺-ATPase inhibitory activity (57). Furthermore, boswellic acid-induced gastroprotection depends on generation of cytoprotective PG, enhanced gastric mucosal resistance, and inhibition of leukotriene synthesis (3). Similarly, *Galipea longiflora* (Rutaceae) protect gastric mucosa by enhancement of mucus content and antisecretory activity (53). In addition to its antioxidant activity, *Cissus sicyoides*, induce increase of NO and SH compounds and enhances defense mechanism (58). In pylorus-ligated rats, beside its antisecretory action, *Momordica charantia* L. extract also significantly increases gastric mucosal content (56). Increase in Glycoprotein level, gastric mucin content and SH concentration are essential for the gastroprotection. Their levels are raised by treatment with *Cissus quadrangularis* extract (49)

7. Herbs and antioxidants

Recently, oxidants are found to play a critical role in PUD pathogenesis. Experimental NSAID and ethanol induced microvascular and gastric mucosal injuries are at least partially caused by ORS release (59). Therefore, implementations of agents with antioxidative properties are useful for the prevention of injuries and promotion of gastric ulcer healing. Many spices have phytochemicals with antioxidative activities. For instance, Coriander contains many antioxidant constituents including d-linalool, borneol, geraniol, geranyl acetate, camphor, carvone, which are responsible for its antioxidative property (60). Black cumin (*Nigella sativa*), piperine and thymoquinone, the active constituents of pepper and *Nigella sativa*, respectively have also the ability to inhibit ROS in experimentally induced gastric lesions in rats (29, 60, 61).

Many other spices/herbs and plant extracts protect against experimentally-induced gastric mucosal injuries through their potential antioxidative effect. These include ginger rhizome, carotenoid and astaxanthin esters, *Cissus sicyoides* extract, isopulegol and the herb collection Korniozil (1, 57, 58, 62, 63), respectively. Through the interaction with endogenous PG and antioxidative properties, isopulegol, monoterpene a constituent of essential oils of several aromatic plants, induce significant gastroprotection. Total carotenoid and astaxanthin esters increase the levels of the antioxidant enzymes catalase, SOD, and GPx in gastric homogenate and protect gastric mucin (57). Korniozil also protects against experimentally induced stress ulcers, with restoration of lipid peroxidation and antioxidative system function along with enhancement of gastric mucous coat regeneration (63). Ginger rhizome extract gastroprotective activity is also based on restoration of antioxidant enzymes and gastric mucin generation in addition to inhibitory effect on *H. pylori* growth (1). Due to its antioxidative properties, *Cissus sicyoides* oral extract increase also NO and SH and induces also protection (58]

The alkaloid indigo, obtained from the leaves of *Indigofera truxillensis* Kunth (Fabaceae), prevents ethanol induced depletion of SH and GPx activity, inhibits GR and MPO activities and partially inhibits gastric mucosa DNA damage caused by ethanol (64).

8. Herbs combined activities

Due the presence of several active constituents, some spices/herbs and plant extracts protect the gastric mucosa via different mechanisms. For instance, Weikang decoction acts as antisecretory, cytoprotective and antioxidative agent. It enhances mucosal thickness, NO in

gastric tissue, PGE₂ in plasma, (EGF) content in gastric juice and SOD in plasma. In addition it inhibits also MDA and endothelin in plasma (65). Many other spices like Rocket *Eruca sativa*, black cumin, black pepper, clove, cardamom, caraway, peppermint, saffron, coriander and anise possess also antisecretory, cytoprotective and antioxidative activities (4, 29, 66-73). They also replenish gastric wall mucus concentration and SH levels and significantly reduce MDA level.

Besides its *H. pylori* bactericidal effect, *Davilla elliptica* also enhances NO, H₂O₂, TNF - α production and GSH bioavailability. These activities are related to its phytochemical constituents acylglycoflavonoids, phenolic acid derivatives and tannins (74). Also Brazilian medicinal plant methanolic extracts have an anti-*H. pylori* effect and protect the gastric mucosa by increasing PGE₂, antisecretory and gastroprotective properties (75) The gastroprotection of *Vochysia tucanorum* Mart. methanolic extract and buthanolic fraction provided by the antioxidant activity and maintenance of gastric mucosa NO levels is interrelated to its phytochemical constituent Triterpenoid (76)

9. Herbs- PG interaction

Gastric protection is maintained in a state of equilibrium between aggressive and protective factors. In experimental ulcer model, indomethacin increases acid secretion, activates oxidative stress and inhibits the release of cyclooxygenase-1 (COX-1), PGE₂, bicarbonate, and mucus (77). Similar to conventional NSAID, COX-2 inhibitors also delay the healing of chronic gastric ulcer and suppress the epithelial cell proliferation, angiogenesis and maturation of the granulation tissue in experimental animals. COX-2 is important for gastric mucosal defense (78). Indomethacin-induced gastric damage is associated with an increase of acid and oxidative parameters and inhibition of protective factors such as COX-1, PGE₂, bicarbonate, and mucus release (77).

Generally, PG are products of arachidonic acid and their biosynthesis is influenced by local, hormonal and neural factors. They stimulate gastric and duodenal bicarbonate secretion and the production of mucus glycoproteins. PG are also able to protect the gastric mucosa against experimentally induced gastric injuries in an acid independent manner known as "cytoprotection" (79). PG play a pivotal role in the gastric mucosal defensive system and contribute to the overall protective process against gastric mucosal injuries. They exhibit a variety of defensive mechanisms including mucus-alkaline secretion, mucosal hydrophobicity, mucosal microcirculation, tissue lysosomes stabilization, SH preservation, rapid proliferation and mucosal cells renewal. Mucosal integrity protection can also be accomplished even by small quantities of PG. Gastroprotection is attained by stimulation of mucosal protective PG biosynthesis or by the inhibition of preulcerogenic arachidonic acid metabolites (80). In addition, PG antiulcer activity is determined mainly by their antioxidant property with inhibition of lipid peroxidation as well as SOD and catalase activities (81).

Other mediators involved in gastric mucosal protection besides PG include growth factors, NO, CGRP and some gut hormones such as gastrin and CCK. In addition, leptin, ghrelin and gastrin-releasing peptide (GRP) have also the ability to protect gastric mucosa against corrosive agents-induced mucosal damage. Gut hormones protective activity is attributed to PG release (41). Ulcer healing is controlled by contribution of growth factors and gut hormones, increase of COX-2 induction and local PGE release in the ulcer area. Endogenous PG generated at ulcer margin play a key role in ulcer cure (82)

In acute injury, in the presence of PG-mediated paracellular space closure, mucosal permeability, PG helps the mucosal permeability to recover with epithelial restitution (83,

84). The ulcer healing by endogenous PG is mediated by PGEP4 receptors, as well as involvement of COX-2 in the early stage and COX-1 in the late stage of healing. Bacterial lipopolysaccharide contributes also through COX and endogenous PG genes activation to gastric mucosal protection in rats (85).

Mainly through their effect on PG, several plant extracts promote ulcer healing. For instance, *Hyptis spicigera* essential oil major constituents, monoterpenes, enhances PGE₂-induced gastric mucus and reduces ulcer size in addition to increasing COX-2 and EGF expression in gastric mucosa and acceleration of ulcer healing (86).

Other spices and plant extracts involved in activation of PG synthesis and healing of gastric mucosal injuries include Boswellic acid, isopulegol, *Teucrium polium* (3, 62, 87). Endogenous PG and PGE₁ receptors play a key role in the adaptive protection (88). Chili is believed to be detrimental to the gastric mucosa, however, its active ingredient capsaicin, decreases acid secretion and activates the defensive system by enhancing mucus, alkali secretions as well as mucosal microcirculation and hence, it prevents ulcer formation. Furthermore, capsaicin stimulates afferent neurons in the stomach and transmits signals to the central nervous system, which trigger an anti-inflammatory response and gastroprotection (89). Furthermore, *Citrus lemon*, *Alchornea triplinervia* and *Myristica malabarica* have demonstrated gastroprotective effect. *Citrus lemon* belongs to Rutaceae family and contains two main components, limonene and β -pinene. In ethanol and indomethacin gastric ulcer models, while *Citrus lemon* and limonene induce complete gastroprotection, β -pinene is not effective. *Citrus lemon* and limonene protective effect is linked with PGE₂ and mediated by enhancing mucus secretion, HSP-70 and VIP (90). The antiulcer effect of ethyl acetate fraction of *Alchornea triplinervia*, a medicinal plant used in Brazil to treat gastrointestinal ulcers, is mainly related to its flavonoids content and mediated by increasing gastric mucosal prostaglandin PGE₂ levels (75).

While PGE₂, and vascular endothelial Growth Factor (VEGF) levels decrease, EGF and endostatin levels increase in indomethacin- induced ulceration in mice. Through modulation of PG synthesis and angiogenesis, *Myristica malabarica* plant extract restores these parameters. In comparison omeprazole, which offered similar healing, did not alter these parameters (91).

Coenzymes Q₁₀, an essential cofactor in the mitochondrial electron transport pathway possess a potent antioxidant action. Pretreatment of indomethacin induced gastropathy with CoQ₁₀ prevents ROS generation, mitochondrial dysfunction, vascular permeability erosions, ulcers and helps to restore PGE₂, NO and GSH levels (92).

10. Herbs and EGF

Growth factors and their receptors are also important for maintaining physiological function of gastric mucosa. They maintain and enhance defensive and inhibit aggressive factors. Following acute mucosal injury and during the initial stages of experimental gastric ulcer healing, R-associated tyrosine kinase is essential for regulation of cell proliferation, EGFR gene activation, EGFR phosphorylation, and increased mitogen-activated protein (MAP) kinase activity. *H. pylori* is a major cause of PUD and contributes also to inhibition of healing. In experimental gastric ulcer model, *H. pylori* vacuolating cytotoxin interferes with ulcer healing and inhibits cell proliferation, binding of EGF to its receptor, EGF-induced EGFR phosphorylation, and MAP and extracellular signal-related kinase (ERK-2) activation (93,94).. Growth factors and their receptors are pivotal for the process of gastroprotection

and ulcer healing. EGF and transforming growth factor (TGF)- α and their common receptor (EGFR) inhibit gastric secretion, boost overexpression of growth factors, blood flow at ulcer margin and promote cell proliferation with ulcer healing (95). The process of gastric mucosal tissue repair and healing is controlled by EGFR activation (96). EGF-induced gastric epithelial cells proliferation is likely intervened by ERK /COX-2 pathway (97). Various GF exhibit different functions of the mucosal repair. They are implicated in the process of tissue healing with cell migration, proliferation, differentiation, secretion, and degradation of extracellular matrix. While EGF, TGF- α , and trefoil factors (TFFs), usually present in the gastric juice or mucosa, as well as hepatocyte growth factor (HGF) are responsible for epithelial structure reconstitution, basic fibroblast growth factor (bFGF), (VEGF), transforming growth factor- β (TGF- β) and platelet derived growth factor (PDGF), are essential for connective tissue reconstitution(98,99).

In gastric mucosal injury, EGF released from salivary glands and TGF- α from gastric mucosa are of particular value in mucosal integrity maintenance and repair. EGF and TGF- α have similar spectra of biological activity in the repair mechanism. Accumulation of EGF and EGFR overexpression in the ulcer area contributes together to repair process. During the ulcer healing process they activate cells migration from the ulcer margin and cell proliferation along with formation of granulation tissue and microvessels, angiogenesis (100). During initial stage of experimental ulcer healing, EGFR-associated tyrosine kinase plays an essential role in the regulation of cell proliferation by activation of the EGFR gene, EGFR phosphorylation, and enhancement of MAP kinase activity. The presence of *H. pylori* vacuolating cytotoxin counteracts this process (93).

Numerous growth factors accelerate gastric epithelial and mesenchymal injury healing in vitro with acceleration of cell migration and proliferation. Gastric epithelial healing is mainly accelerated by a group of growth factors including EGF, TGF- α and HGF, while mesenchymal healing is predominantly accelerated by TGF- β and bFGF. Both, gastric epithelial and mesenchymal injury healing are significantly accelerated by PDGF, factor-betabeta and insulin-like growth factor-1 (IGF-1). During the healing process, IGF-1 regulates the gastric epithelial-mesenchymal interaction (96,101).

In injured gastric mucosa, growth factors TGF- α , HGF and IGF accelerate epithelial restitution and variably regulate the regeneration of human gastric epithelial cells through modulation of cell shape adaptation, migration and proliferation (102). Growth factors endorse EGFR-dependent PI3K activation, which promotes cell migration and restitution in injured human gastric epithelial monolayers (103).

Smoking is known as a risk factor for PUD. The detrimental effect of smoking is exhibited by inhibition of cell proliferation, mucus secretion and angiogenesis due to deficiency in EGF biosynthesis and its mRNA expression. The shortage of these factors is responsible for the delay in ulcer healing (104).

Several spices and plant extracts such as Mexican tea herb and pilular adina herb, Chuanxiong spices, Capsaicin, Kuyiangping, Weitongning and Angelica sinensis interact with EGF synthesis and hence contribute to the gastroprotection. For instance, Mexican tea herb and pilular adina herb stimulate NO, EGF secretion and EGFR expression and herewith protect the gastric mucosa integrity (105). Capsaicin-sensitive nerves induced ulcer healing is mediated by stimulation of EGF expression in salivary glands, serum and gastric mucosa (106). Also Kuyiangping, promotes ulcer healing in rats and decreases recurrence via increased expression of EGF and EGFR mRNA (107). Furthermore, Weitongning herb increases EGF and NO content in ulcer scars, and hence improves ulcer healing and reduces recurrence (108). In experimental myocardial infarction, Angelica and Chuanxiong spices

promote endothelial cell proliferation and VEGF expression(109) and likewise may also promote angiogenesis and tissue repair in experimental ulcer. In indomethacin-induced gastric mucosal injury, crude extract from *Angelica sinensis* promotes EGF-mediated gastric mucosal healing via DNA synthesis, stimulation and augmentation of EGF mRNA expression (110). *Picrorhiza kurroa* (Scrofulariaceae) rhizomes possess an antioxidative property indicated by reduction of thiobarbituric acid reactive substances (TBARS) and protein carbonyl in addition to enhancing expression of EGF, VEGF, COX-1 and 2 enzymes associated with an increase of mucin and mucosal PGE₂, which explain its ability to heal indomethacin-induced acute gastric injury in mice (111). Similarly, *Myristica malabarica* spice constituting two major antioxidants, malabaricone B and malabaricone C suppressed thiobarbituric acid reactive substances and protein carbonyls levels. Malabaricone C is more potent in modulating expression of EGF receptor and COX isoforms, mucin secretion, PGE₂ synthesis and in controlling all these factors (112). Furthermore, ulcer cure by malabaricone B and malabaricone C is related to their ability to modulate angionetic factors. They significantly increase the mucosal EGF level serum VEGF level and microvessels formation. In contrary, the healing effect of misopristol and omeprazole is not correlated with angiogenesis enhancement (113).

11. Herbs and nitric oxide

In combination with other factors, NO significantly add to mucosal protection. The inflammatory process is mediated by inducible nitric oxide synthase (iNOS) and interleukin-8 (IL-8). Nitric oxide donors (SIN-1 and NOC-18) augment IL-8 and nitrite in mRNA, expression of IL-8. Production of large amounts of NO by iNOS may activate NF-kappaB and AP-1 and the expression of IL-8 in gastric epithelial cells (114). While iNOS is found in inflammatory cells in ulcer bed, NOS is located at the vascular endothelium and mucosal cells in normal and ulcerated gastric tissues. Endothelial NOS and NO significantly contribute to ulcer healing (106,115). Maintenance of NO synthesis is essential for an adequate mucosal defense. Conversely, Inhibition of NO synthesis in mucosal injury models is associated with an increase in ulcer index and asymmetric dimethylarginine (ADMA) levels along with a significantly decreased dimethylarginine dimethylaminohydrolase (DDAH) activity. ADMA Administration is associated with an inflammatory process with inhibition of NO synthesis and elevation of TNF- α levels and indicates the importance of ADMA in precipitating gastric mucosal injury (116). Such a process can be prevented by the use of extracts obtained from herbs and plants rich in phenolic compounds. Methanolic extract and buthanolic fraction of *Vochysia tucanorum* Mart., possess an antioxidant activity and protect NO levels in gastric mucosa. This protective effect is probably mediated by its phenolic compounds containing various active phytochemical constituents, triterpenoids (76). Triterpenoids are also active constituents of *Croton reflexifolius* and may explain its gastroprotective effect. Pretreatment with NOS inhibitor attenuates the gastroprotective effect induced by polyalthic acid (117). Furthermore, plant-extract-induced gastroprotective activity is likely related to the enhancing effect on release of NO in addition to NOS inhibitor expression and gastric microcirculation (118).

12. Herbs and SH compounds

The pathogenesis of gastric ulcer is complex. Several endogenous substances including SH compounds are important for the cytoprotection. They are involved in motivation of PG

synthesis, protection of gastric mucosal integrity as well as in the antioxidative process. SH mucosal concentration is suppressed, especially in ethanol-induced gastric mucosal injuries. Preservation of mucosal microcirculation for rapid restitution and cell proliferation is considered as a key target of gastroprotection by either PG or SH compounds (119).

Several spices and plant extracts have protective effect against ethanol-induced SH depletion. Among these spices, Black seed, coriander, peppermint, black pepper, clove, anise aqueous suspension, and rocket replenish ethanol-induced gastric wall mucus and SH depletion in experimental studies (4,66,67,71-73). Similarly, Ginkgo biloba extract preserves mucosal function via inhibition of ethanol-induced SH and gastric wall mucus depletion and lipid peroxidation (120). Methanolic extracts of *C. sicyoides* and *Commiphora opobalsamum* (L.) Engl. (Balessan) also enhances the defense system in rodents and inhibits gastric injuries through SH and NO involvement (58).

13. Herbs and cytokines

Altered immune system function significantly contributes to the pathogenesis of ulcer disease, particularly T-helper lymphocytes and released cytokines. The gastroprotection induced by *Phyllanthus emblica* L. also upregulates anti-inflammatory cytokine IL-10 concentration through its antioxidative activity, modulates anti-inflammatory cytokines and inhibits pro-inflammatory cytokines TNF- α and IL-1 β (121).

The process of ulcer formation is considerably induced and regulated by IL-1 β , TNF- α , IL-4, -6, -8, -12 cytokines. Cytokines, IL-1 β and IL-1RN genes modulate the inflammatory response and therefore play an important role in the course of the disease (122).

In many gastric injuries, TNF- α is involved in the induction of chemokine expression. It increases the number of macrophages and monocyte chemoattractant protein-1 (MCP-1) mRNA expression in mucosal scar. Increased MCP-1 may play a key role in regulating leukocyte recruitment and chemokine expression in gastric ulcer. TNF- α increases also macrophage inflammatory protein (MIP)-2 and cytokine-induced neutrophil chemoattractant (CINC-2 α) mRNA expression and MPO activity (123). Cytokine gene polymorphisms influence mucosal cytokine expression and the degree of inflammation in *H. pylori* infection (124).

Furthermore, IL-1 β enhances adhesion molecules expression, intercellular adhesion molecule 1 and leucocytic β 2 integrins as well as the concentrations of TNF- α in ulcer scar and contributes to the recurrence of gastric ulcers in rats. The presence of gastric acid is important for the recurrence process of IL-1 β -induced gastric ulcer. Gastric acid activates the inflammatory process in scarred mucosa during ulcer recurrence (125).

The outcome of *H. pylori* infection is influenced by the host response, which in susceptible individuals determines the development of ulcer. In *H. pylori* infected antral mucosa response is associated with an increase of proinflammatory IL-1 β , IL-6, TNF α cytokines, and IL-8; the immunoregulatory gamma interferon (IFN- γ); and the anti-inflammatory TGF- β (126). A correlation between genetic polymorphisms and *H. pylori*-related diseases is well-established. While IFN- γ +874 AA genotype is associated with *cagA* positive infections, IL-10 -819 TT and TNF-A -857 TT are associated with intestinal metaplasia and duodenal ulcer, respectively (127).

Among various gastropathies gastritis is the only gastric disorder associated with significant oxidative stress marker expression of TNF- α , IL-8 and *H. pylori cagA*+/*vacAs1* genotype. These probably represent the main oxidative markers responsible for ROS level increase with a decrease of the expression of the Manganese superoxide dismutase (MnSOD) and GPx (128).

In Western countries, polymorphism of pro-inflammatory cytokine genes is associated with the development of duodenal ulcer and gastric cancer. Similarly, polymorphisms in TNF- α rather than IL-1 β are associated with an increased risk for gastric ulcers and gastric cancer in Japan. Increased risk of gastric ulcer development is associated with carriage of the alleles TNF- α -857 T, TNF- α -863 A and TNF- α -1031 C. Simultaneous carriage of more than one high-producer allele of TNF- α further increase the risks for gastric ulcer and cancer (129). In chronic *H. pylori* infection Pro-inflammatory cytokines are produced in the gastric mucosa by inflammatory cells. In contrast to Asians, in western population the inflammatory cytokine gene polymorphisms IL-4-590, IL-6-572 and IL-8-251 are more associated with development of PUD. Polymorphisms of these and other cytokines such as IL-1 β , IL-1RN and TNF- α , may help to predict those at higher risk to develop peptic ulcer and those, who require *H. pylori* eradication(130).

Spices and other plant extracts may interfere with cytokines function, regulate the inflammatory process and help in ulcer healing. In gastric ulcer model, both curcumin and bisdemethoxycurcumin, a yellow pigment in rhizomes of *Curcuma longa*, promote gastric ulcer healing. While curcumin suppress iNOS and TNF- α protein production, bisdemethoxycurcumin lowers the increased iNOS protein expression level without any effect on TNF- α . The gastroprotective property of bisdemethoxycurcumin is related to its capability to decrease gastric acid secretion and suppress iNOS-mediated inflammation (131). Medicinal plants may also modulate lipopolysaccharide-induced proinflammatory cytokine production in murine macrophage cells and in mice treated with the stimulant lipopolysaccharide. This has been demonstrated by the use of three herbal constituents, apigenin (chamomile), ginsenoside Rb1 (ginseng) and parthenolide (feverfew). All of these herbal constituents have inhibited lipopolysaccharide-induced IL-6 and/or TNF- α production in culture(132).

14. Herbs and *H. pylori*

H. pylori represents the main cause for PUD and its eradication is imperative for ulcer healing and reduction of ulcer recurrence rate. The current eradication rate is below 90% and the resistance rate is growing up. Therefore, the search for potent *H. pylori* bactericidal agents from plants resources is emerging. Several spices and plant extracts possess *H. pylori* growth inhibitory activities. Curcumin (133), black cumin (134), eugenol, cinamaldehyde (135), turmeric, cumin, ginger, chilli, borage, black caraway, oregano and parsley (136) have an anti-*H. pylori* activity. Oil extract of *Chamomilla recutita* affects *H. pylori* morphological and fermentative properties and inhibits urease production (137). *H. pylori* adhesion to the gastric mucosa, an important stage of infection is inhibited by extracts of turmeric, borage and parsley (136), curcumin and its methanolic extract restrain the growth of all strains of *H. pylori* in vitro (133)]. Moreover, eugenol and cinnamaldehyde have prevented growth of *H. pylori* obtained from human gastric tissue, and inhibited the growth of all 30 tested *H. pylori* strains, with a lack of resistance (135). Besides, phenolic compounds of Oregano (*Origanum vulgare* L.), a Mediterranean herb, possess an inhibitory effect on *H. pylori* growth (138).

Also, aqueous-ethanol extracts of over 25 of Pakistani medicinal plants including *Mal. philippines* (Lam) Muell. *Mallotus philippines* (Lam) Muell., *Curcuma amada* Roxb., *Myristica fragrans* Houtt., and *Psoralea corylifolia* L have potent anti-*H. pylori* activity (139). In addition, methanolic extract of 25 of 50 Taiwanese folk medicinal plants have also

demonstrated compelling anti-*H. pylori* action (140). Furthermore, of 53 Mexican traditional medicinal plants especially extracts of *Artemisia ludoviciana* subsp. *mexicana*, *Cuphea aequipetala*, *Ludwigia repens*, and *Mentha x piperita* and methanolic extracts of *Persea americana*, *Annona cherimola*, *Guaiacum coulteri*, and *Moussonia deppeana* have verified a persuasive *H. pylori* inhibitory effect (141).

At last, the anti- *H. pylori* effect of 70 Greek plant extracts and a variety of commercially available herbs used in traditional medicine such as extracts of *Chamomilla recutita*, *Conyza albida*, *Origanum vulgare* *Anthemis melanolepis*, *Cerastium candidissimum*, *Dittrichia viscosa*, and *Stachys alopecuroides* have inhibited a standard strain and 15 *H. pylori* clinical isolates (142).

15. Adverse events

Spices, herbs and other plant extracts have been used in traditional medicine for thousands of years. Recently, in several parts of the world there is a growing acceptance for using these agents to treat various conditions including PUD. Most of these extracts have been effective; however their safety and toxicity have not been well-evaluated. The increasing use of herbal medicine is expected to be more frequently associated with adverse reactions. Clinical evaluation of these adverse effects is not easy due lack of standardization, randomization, adequate number of patients and difficulty in using an appropriate placebo. Herbs are believed to be safe and have no adverse effect. However similar to other drugs they may induce intrinsic or extrinsic adverse effects. Some of their multiple constituents, such as anti-cancer plant-derived drugs, digitalis and the pyrrolizidine alkaloids are cytotoxic. Nevertheless, their adverse effects are less frequent than those of synthetic drugs (143).

Hepatotoxicity induced by curcumin and its derivatives (144) as well as by turmeric and its ethanolic extract in vulnerable mice has been reported (145). Also animals treated with Cinnamon *zeylanicum*, *Piper longum* and *R. chalepensis* have developed abnormalities in liver, spleen, lung or reproductive organs, in addition to an increase in count and motility of sperm and decrease in hemoglobin level (146,147). Kava (*Piper methysticum*), used as anxiolytic herb in Western countries has been potentially found to be hepatotoxic. Its hepatotoxicity is correlated with overdose, prolonged treatment, concurrent medication, and the quality of raw material (148). Suspected herb-induced liver injury (HILI) is evaluated by the causality score using this multidisciplinary approach and Roussel Uclaf Causality Assessment Method (RUCAM) (149).

In addition to hepatic toxicity, alteration of body weight has been described in rodents treated with *Foeniculum vulgare* ethanolic extracts and *Ruta chalepensis* (150). Furthermore, in experimental model, piperine has decreased mating performance and fertility and intrauterine injection has caused loss of implants without histological abnormalities (151). Herbal-induced toxicity is influenced by herbs related factors (quality, dose and nature of constituents) and individual risk factors (genetics, age, concomitant drugs, and concomitant diseases) (152). Therefore, simultaneous administration of herbs with conventional medications should generally be discouraged (153).

Herbal medicine-associated adverse reactions are expected to occur more frequently as a result of the fast mounting use of these agents in treatment. Some commonly used herbs like St John's wort (*Hypericum perforatum*), a popular herbal anti-depressant, lead to a decrease of the activity of immunosuppressive agents i.e. cyclosporine and subsequent tissue rejection in transplanted patients. Like other medicinal plants it also interferes with cytochrome P450 activity and metabolism of other drugs (154).

Examples of Drugs known to interact with St John's wort include besides cyclosporine tacrolimus as well as HIV non-nucleoside and protease inhibitors (155). Other drugs interfering with St. John's wort CYP 3A4 induction include , oral contraceptives.and indinavir(156).

Literature review of 128 case reports or case series, and 80 clinical trials have revealed that St John's wort-induced cytochrome P450 and P-glycoprotein induction, decreases plasma levels of a large variety and frequently used medications. Clearance of caffeine and midazolam may be influenced by Echinacea (157).

Herbal agents such as St. John's wort, interact differently with various drugs. It may increase the clearance of some medications via cytochrome P-450 mixed-function oxidase or through P-glycoprotein efflux pump modulation. On the other hand, it may decrease digoxin, theophylline, warfarin, protease inhibitors, cyclosporine, tacrolimus, and tricyclic antidepressants concentration with subsequent reduction of their therapeutic effect. A third category of drugs such as procainamide carbamazepine and mycophenolic acid are not affected by St. John's wort. herb (158).

Therapeutic drug monitoring is usually estimated by immunoassay technique. The potential interference St. John's wort, with commonly by this method monitored drugs has been evaluated. A significant interference with digoxin, quinidine, procainamide, N-acetyl procainamide theophylline, tricyclic antidepressants, phenytoin, carbamazepine, valproic acid and phenobarbital serum levels is lacking (159). Due to unwanted effects, ginseng and ginkgo should not be combined with anticoagulants and valerian with barbiturates (160). Elderly patients are more likely to develop diseases and ingest more medications. They are also prone to develop suppression of cytochrome P450 (CYP) activity. Taking herbal agents make them more vulnerable to herb-drug interactions (161). Herbal toxicity may also affect other central organs like the kidney. Case reports of interstitial fibrosis progressing to chronic renal failure and termed as aristolochic acid nephropathy may complicate treatment with slimming herbs belonging to Aristolochia family (162). Despite all of these reports of adverse events, spices are generally safe when used in standard doses. Popular traditional Chinese medicine has relatively less adverse effects and appears safer than other drugs (163).

The safety of herbal agents during pregnancy has been evaluated in 392 pregnant women 8% have reported taking chamomile, licorice, fennel, aloe, valerian, Echinacea oil 27, propolis and cranberry. Only four out 109 have reported insignificant adverse events in form of constipation after tisane, rash and itching after local application of aloe or almond oil. A higher incidence of threatening miscarriage and preterm labors was observed among regular users of chamomile and licorice (164).

In disparity, many spices and plant extracts, in commonly used dose, up to 500mg/kg body weight have not exhibited adverse effect. These include cardamom (62, 68), black pepper (66), clove (67), caraway (69), saffron (70), coriander(71), peppermint (72), anise (73), davilla elliptica and nitida (74) ,Brazilian medical plants (75) and Alchornea triplinervia (76) and Hyptis spicigera Lam (86). Even in pregnancy, ginger, peppermint, and Cannabis have been used to treat nausea were effective and lack clinical evidence of harm (165). Clinically, spices like turmeric and curcumin have been well-tolerated even with high doses and lack any toxicity (166).

16. References

- [1] Nanjundaiah SM, Annaiah HN, M Dharmesh S. Gastroprotective Effect of Ginger Rhizome (*Zingiber officinale*) Extract: Role of Gallic Acid and Cinnamic Acid in

- H⁺, K⁺-ATPase/H. pylori Inhibition and Anti-oxidative Mechanism. *Evid Based Complement Alternat Med.* 2009; 1-13.
- [2] Choi SM, Shin JH, Kang KK, Ahn BO, Yoo M. Gastroprotective effects of DA-6034, a new flavonoid derivative, in various gastric mucosal damage models. *Dig Dis Sci* 2007;52: 3075–3080. [PubMed]
- [3] Singh S, Khajuria A, Taneja SC, Khajuria RK, Singh J, Johri RK, Qazi GN. The gastric ulcer protective effect of boswellic acids, a leukotriene inhibitor from *Boswellia serrata*, in rats. *Phytomedicine* 2008; 15:408–415. [PubMed]
- [4] Alqasoumi S, Al-Sohaibani M, Al-Howiriny T, Al-Yahya M, Rafatullah S. Rocket “*Eruca sativa*”: a salad herb with potential gastric anti-ulcer activity. *World J Gastroenterol* 2009;15:1958–1965. [PubMed]
- [5] Chernomorets NN, Seleznev AV, Revutskii BI, Alifanova RE, Kravchenko ZV, Cherkasskaia EP. [The differentiated phytotherapy of patients with duodenal peptic ulcer] *Lik Sprava* 1992:112–115. [PubMed]
- [6] Chakürski I, Matev M, Stefanov G, Koichev A, Angelova I. [Treatment of duodenal ulcers and gastroduodenitis with a herbal combination of *Symphitum officinalis* and *Calendula officinalis* with and without antacids] *Vutr Boles* 1981;20:44–47.
- [7] Al-Howiriny T, Al-Sohaibani M, Al-Said M, Al-Yahya M, El-Tahir K, Rafatullah S. Effect of *Commiphora opobalsamum* (L.) Engl. (Balessan) on experimental gastric ulcers and secretion in rats. *J Ethnopharmacol* 2005;98:287–294. [PubMed]
- [8] Schiestl RH, Chan WS, Gietz RD, Mehta RD, Hastings PJ. Safrole, eugenol and methyleugenol induce intrachromosomal recombination in yeast. *Mutat Res* 1989;224:427–436. [PubMed]
- [9] Lo YC, Yang YC, Wu IC, Kuo FC, Liu CM, Wang HW, Kuo CH, Wu JY, Wu DC. Capsaicin-induced cell death in a human gastric adenocarcinoma cell line. *World J Gastroenterol* 2005;11:6254–6257. [PubMed]
- [10] Mothana RA, Gruenert R, Bednarski PJ, Lindequist U. Evaluation of the in vitro anticancer, antimicrobial and antioxidant activities of some Yemeni plants used in folk medicine. *Pharmazie* 2009;64:260–268. [PubMed]
- [11] Lichtenberger LM, Romero JJ, Carryl OR, Illich PA, Walters ET. Effect of pepper and bismuth subsalicylate on gastric pain and surface hydrophobicity in the rat. *Aliment Pharmacol Ther* 1998;12:483–490. [PubMed]
- [12] Al Mofleh IA, Alhaider AA, Mossa JS, Al-Sohaibani MO, Al-Yahya MA, Rafatullah S, Shaik SA. Gastroprotective effect of an aqueous suspension of black cumin *Nigella sativa* on necrotizing agents-induced gastric injury in experimental animals. *Saudi J Gastroenterol* 2008;14:128–134. [PubMed]
- [13] Pan JS, He SZ, Xu HZ, Zhan XJ, Yang XN, Xiao HM, Shi HX, Ren JL. Oxidative stress disturbs energy metabolism of mitochondria in ethanol-induced gastric mucosa injury. *World J Gastroenterol* 2008 ; 14;14 :5857-5867.
- [14] Hirokawa M, Miura S, Yoshida H, Kurose I, Shigematsu T, Hokari R, Higuchi H, Watanabe N, Yokoyama Y, Kimura H, Kato S, Ishii H. Oxidative stress and mitochondrial damage precedes gastric mucosal cell death induced by ethanol administration. *Alcohol Clin Exp Res* 1998 ;22(3 Suppl):111S-114S.
- [15] Li NS, Luo XJ, Zhang YS, He L, Liu YZ, Peng J. Phloroglucinol protects gastric mucosa against ethanol-induced injury through regulating myeloperoxidase and catalase activities. *Fundam Clin Pharmacol.* 2010. [Epub ahead of print] PMID: 20880383)

- [16] Farias-Silva E, Cola M, Calvo TR, Barbastefano V, Ferreira AL, De Paula Michelatto D, Alves de Almeida AC, Hiruma-Lima CA, Vilegas W, Brito AR. Antioxidant activity of indigo and its preventive effect against ethanol-induced DNA damage in rat gastric mucosa. *Planta Med* 2007 ;73 :1241-1246.
- [17] Liu CS, Cham TM, Yang CH, Chang HW, Chen CH, Chuang LY. Antibacterial properties of Chinese herbal medicines against nosocomial antibiotic resistant strains of *Pseudomonas aeruginosa* in Taiwan. *Am J Chin Med* 2007;35:1047-1060. [PubMed]
- [18] Seneviratne CJ, Wong RW, Samaranyake LP. Potent anti-microbial activity of traditional Chinese medicine herbs against *Candida* species. *Mycoses* 2008;51:30-34. [PubMed]
- [19] Hitokoto H, Morozumi S, Wauke T, Sakai S, Kurata H. Inhibitory effects of spices on growth and toxin production of toxigenic fungi. *Appl Environ Microbiol* 1980;39:818-822. [PubMed]
- [20] Karapinar M. Inhibitory effects of anethole and eugenol on the growth and toxin production of *Aspergillus parasiticus*. *Int J Food Microbiol* 1990;10:193-199. [PubMed]
- [21] El Garhy MF, Mahmoud LH. Anthelmintic efficacy of traditional herbs on *Ascaris lumbricoides*. *J Egypt Soc Parasitol* 2002;32:893-900. [PubMed]
- [22] Kiuchi F, Goto Y, Sugimoto N, Akao N, Kondo K, Tsuda Y. Nematocidal activity of turmeric: synergistic action of curcuminoids. *Chem Pharm Bull (Tokyo)* 1993;41:1640-1643. [PubMed]
- [23] Sharma JN, Srivastava KC, Gan EK. Suppressive effects of eugenol and ginger oil on arthritic rats. *Pharmacology*. 1994;49:314-318. [PubMed]
- [24] Li EK, Tam LS, Wong CK, Li WC, Lam CW, Wachtel-Galor S, Benzie IF, Bao YX, Leung PC, Tomlinson B. Safety and efficacy of *Ganoderma lucidum* (lingzhi) and San Miao San supplementation in patients with rheumatoid arthritis: a double-blind, randomized, placebo-controlled pilot trial. *Arthritis Rheum*. 2007;57: 1143-1150. [PubMed]
- [25] Spiller F, Alves MK, Vieira SM, Carvalho TA, Leite CE, Lunardelli A, Poloni JA, Cunha FQ, de Oliveira JR. Anti-inflammatory effects of red pepper (*Capsicum baccatum*) on carrageenan- and antigen-induced inflammation. *J Pharm Pharmacol* 2008;60: 473-478. [PubMed]
- [26] Liu J. Pharmacology of oleanolic acid and ursolic acid. *J Ethnopharmacol* 1995;49 :57-68.
- [27] Morita T, Jinno K, Kawagishi H, Arimoto Y, Sukanuma H, Inakuma T, Sugiyama K. Hepatoprotective effect of myristicin from nutmeg (*Myristica fragrans*) on lipopolysaccharide/d-galactosamine-induced liver injury. *J Agric Food Chem* 2003 ;51:1560-1565.
- [28] Sharma S, Kulkarni SK, Chopra K. Curcumin, the active principle of turmeric (*Curcuma longa*), ameliorates diabetic nephropathy in rats. *Clin Exp Pharmacol Physiol*. 2006 ;33 : 940-945.
- [29] Wongpa S, Himakoun L, Soontornchai S, Temcharoen P. Antimutagenic effects of piperine on cyclophosphamide-induced chromosome aberrations in rat bone marrow cells. *Asian Pac J Cancer Prev* 2007 ;8 : 623-627.
- [30] Oyagbemi AA, Saba AB, Azeez OI. Molecular targets of [6]-gingerol: Its potential roles in cancer chemoprevention. *Biofactors* 2010 ;36 : 169-178.

- [31] Al-Amin ZM, Thomson M, Al-Qattan KK, Peltonen-Shalaby R, Ali M. Anti-diabetic and hypolipidaemic properties of ginger (*Zingiber officinale*) in streptozotocin-induced diabetic rats. *Br J Nutr* 2006;96: 660-666. [PubMed]
- [32] Ejaz A, Wu D, Kwan P, Meydani M. Curcumin inhibits adipogenesis in 3T3-L1 adipocytes and angiogenesis and obesity in C57/BL mice. *J Nutr* 2009;139: 919-925. [PubMed]
- [33] Alwi I, Santoso T, Suyono S, Sutrisna B, Suyatna FD, Kresno SB, Ernie S. The effect of curcumin on lipid level in patients with acute coronary syndrome. *Acta Med Indones* 2008;40: 201-210. [PubMed]
- [34] Alizadeh-Navaei R, Roozbeh F, Saravi M, Pouramir M, Jalali F, Moghadamnia AA. Investigation of the effect of ginger on the lipid levels. A double blind controlled clinical trial. *Saudi Med J* 2008;29: 1280-1284. [PubMed]
- [35] Al-Yahya MA, Rafatullah S, Mossa JS, Ageel AM, Parmar NS, Tariq M. Gastroprotective activity of ginger *zingiber officinale* rosc., in albino rats. *Am J Chin Med* 1989;17: 51-56. [PubMed]
- [36] Rafatullah S, Tariq M, Al-Yahya MA, Mossa JS, Ageel AM. Evaluation of turmeric (*Curcuma longa*) for gastric and duodenal antiulcer activity in rats. *J Ethnopharmacol* 1990;29: 25-34. [PubMed]
- [37] Schmeda-Hirschmann G, Yesilada E. Traditional medicine and gastroprotective crude drugs. *J Ethnopharmacol* 2005 22;100 : 61-66.
- [38] Graham DY, Smith JL, Opekun AR. Spicy food and the stomach. Evaluation by videoendoscopy. *JAMA* 1988;260: 3473-3475. [PubMed]
- [39] Kivilaakso E. Pathogenetic mechanisms in experimental gastric stress ulceration. *Scand J Gastroenterol Suppl* 1985;110: 57-62. [PubMed]
- [40] Nayeb-Hashemi H, Kaunitz JD. Gastroduodenal mucosal defense. *Curr Opin Gastroenterol* 2009;25: 537-543. [PubMed]
- [41] Brzozowski T, Konturek PC, Konturek SJ, Brzozowska I, Pawlik T. Role of prostaglandins in gastroprotection and gastric adaptation. *J Physiol Pharmacol* 2005;56 Suppl 5: 33-55.
- [42] Luo JC, Shin VY, Yang YH, Wu WK, Ye YN, So WH, Chang FY, Cho CH. Tumor necrosis factor-alpha stimulates gastric epithelial cell proliferation. *Am J Physiol Gastrointest Liver Physiol* 2005;288: G32-G38. [PubMed]
- [43] Beales IL. Gastrin and interleukin-1beta stimulate growth factor secretion from cultured rabbit gastric parietal cells. *Life Sci* 2004;75: 2983-2995. [PubMed]
- [44] Borelli F, Izzo AA. The plant kingdom as a source of anti-ulcer remedies *Phytother Res* 2000;14: 581-591.
- [45] A. Jamal, Kalim Javed, M. Aslam, M.A. Jafri. Gastroprotective effect of cardamom, *Elettaria cardamomum* Maton. fruits in rats. *J Ethnopharmacol* 2006 :103, : 149-153.
- [46] Badreldin H Ali, Gerald Blunden, Musbah O Tanira, Abderrahim Nemmar Some phytochemical, pharmacological and toxicological properties of ginger (*Zingiber officinale* Roscoe): a review of recent research. *Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association* 03/2008; 46 : 409-420.

- [47] Eswaran MB, Surendran S, Vijayakumar M, Ojha SK, Rawat AK, Rao ChVG. Gastroprotective activity of *Cinnamomum tamala* leaves on experimental gastric ulcers in rats. *J Ethnopharmacol.* 2010 ;128 : 537-540. [PubMed]
- [48] Al Mofleh IA. Spices, herbal xenobiotics and the stomach: friends or foes? *World J Gastroenterol* 2010 ;16 : 2710-2719. Editorial [PubMed]
- [49] Jainu M, Vijai Mohan K, Shyamala Devi CS. Gastroprotective effect of *Cissus quadrangularis* extract in rats with experimentally induced ulcer. *Indian J Med Res* 2006;123: 799-806. [PubMed] .
- [50] Baggio CH, Freitas CS, Otofujii Gde M, Cipriani TR, Souza LM, Sasaki GL, Iacomini M, Marques MC, Mesia-Vela S. Flavonoid-rich fraction of *Maytenus ilicifolia* Mart. ex. Reiss protects the gastric mucosa of rodents through inhibition of both H⁺,K⁺ - ATPase activity and formation of nitric oxide. *J Ethnopharmacol* 2007;113: 433-440. [PubMed]
- [51] Baek SW, Kim NK, Jin HJ, Koh CW, Kim CK, Kwon OH, Kim JS, Cho MH, Park CK. Anti-ulcer actions of phytosphingosine hydrochloride in different experimental rat ulcer models. *Arzneimittelforschung* 2005;55: 461-465. [PubMed]
- [52] Souccar C, Cysneiros RM, Tanae MM, Torres LM, Lima-Landman MT, Lapa AJ. Inhibition of gastric acid secretion by a standardized aqueous extract of *Cecropia glaziovii* Sneth and underlying mechanism. *Phytomedicine* 2008;15: 462-469. [PubMed]
- [53] Zanatta F, Gandolfi RB, Lemos M, Ticona JC, Gimenez A, Clasen BK, Cechinel Filho V, de Andrade SF. Gastroprotective activity of alkaloid extract and 2-phenylquinoline obtained from the bark of *Galipea longiflora* Krause (Rutaceae) *Chem Biol Interact* 2009;180: 312-317. [PubMed]
- [54] Olaleye SB, Owoyele VB, Odukanmi AO. Antiulcer and gastric antisecretory effects of *Landolphia owariensis* extracts in rats. *Niger J Physiol Sci* 2008;23: 23-26. [PubMed]
- [55] Baggio CH, Freitas CS, Otofujii Gde M, Cipriani TR, Souza LM, Sasaki GL, Iacomini M, Marques MC, Mesia-Vela S. Flavonoid-rich fraction of *Maytenus ilicifolia* Mart. ex. Reiss protects the gastric mucosa of rodents through inhibition of both H⁺,K⁺ - ATPase activity and formation of nitric oxide. *J Ethnopharmacol* 2007;113 : 433-440.
- [56] Alam S, Asad M, Asdaq SM, Prasad VS. Antiulcer activity of methanolic extract of *Momordica charantia* L. in rats. *J Ethnopharmacol* 2009;123 : 464-469.
- [57] Kamath BS, Srikanta BM, Dharmesh SM, Sarada R, Ravishankar GA. Ulcer preventive and antioxidative properties of astaxanthin from *Haematococcus pluvialis*. *Eur J Pharmacol* 2008;590: 387-395. [PubMed]
- [58] de Paula Ferreira M, Nishijima CM, Seito LN, Dokkedal AL, Lopes-Ferreira M, Di Stasi LC, Vilegas W, Hiruma-Lima CA. Gastroprotective effect of *Cissus sicyoides* (Vitaceae): involvement of microcirculation, endogenous sulfhydryls and nitric oxide. *J Ethnopharmacol* 2008 ;117 : 170-174.
- [59] Naito Y, Yoshikawa T. Oxidative stress involvement and gene expression in indomethacin-induced gastropathy. *Redox Rep* 2006;11: 243-253. [PubMed]
- [60] Srinivasan K. Black pepper and its pungent principle-piperine: a review of diverse physiological effects. *Crit Rev Food Sci Nutr* 2007;47: 735-748. [PubMed]

- [61] Kanter M, Demir H, Karakaya C, Ozbek H. Gastroprotective activity of *Nigella sativa* L oil and its constituent, thymoquinone against acute alcohol-induced gastric mucosal injury in rats. *World J Gastroenterol* 2005;11: 6662-6666. [PubMed]
- [62] Silva MI, Moura BA, Neto MR, Tomé Ada R, Rocha NF, de Carvalho AM, Macêdo DS, Vasconcelos SM, de Sousa DP, Viana GS, et al. Gastroprotective activity of isopulegol on experimentally induced gastric lesions in mice: investigation of possible mechanisms of action. *Naunyn Schmiedebergs Arch Pharmacol* 2009;380: 233-245. [PubMed]
- [63] Bogdarin IuA, Potekhin PP, Kozlov DV, Shirokova NIu. [Efficacy of the new collection of herbs at stressful experimental sharp ulcer defects of the gastroduodenal zone] *Eksp Klin Gastroenterol* 2005: 74-78, 102. [PubMed]
- [64] Cola M, Calvo TR, Barbastefano V, Ferreira AL, De Paula Michelatto D, Alves de Almeida AC, Hiruma-Lima CA, Vilegas W, Brito AR. Antioxidant activity of indigo and its preventive effect against ethanol-induced DNA damage in rat gastric mucosa. *Planta Med* 2007 ;73: 1241-1246. [PubMed]
- [65] Fan TY, Feng QQ, Jia CR, Fan Q, Li CA, Bai XL. Protective effect of Weikang decoction and partial ingredients on model rat with gastric mucosa ulcer. *World J Gastroenterol* 2005;11: 1204-1209. [PubMed]
- [66] I.A. Al-Mofleh, A.A. Alhaider, J.S. Mossa, M.O. Al-Sohaibani, S. Rafatullah, S. Qureshi. Inhibition of Gastric Mucosal Damage by *Piper Nigrum* (Black pepper) Pretreatment in Wistar Albino Rats. *PHCOG MAG* 2005; 1 : 64-68.
- [67] I.A. Al-Mofleh, A.A. Alhaider, J.S. Mossa, M.O. Al-Sohaibani, S. Qureshi, S. Rafatullah. Pharmacological Studies on 'Clove' *Eugenia caryophyllata*. *PHCOG MAG* 2005; 1 : 105-109.
- [68] Alhaider A.A., Al-Mofleh I.A., Mossa J.S., Al-Sohaibani M.O., Qureshi S. and Rafatullah S. Pharmacological and Safety Evaluation Studies on "Cardamon" *Elettaria cardamomum*: An Important Ingredient of Gahwa (Arabian Coffee). *Arab Journal of Pharmaceutical Sciences* 2005; 3 : 47-58.
- [69] Alhaider, A.A., I.A. Al-Mofleh, J.S. Mossa, M.O. Al-Sohaibani, S. Rafatullah and S. Qureshi, 2006. Effect of *Carum carvi* on experimentally induced gastric mucosal damage in wistar albino rats. *Int. J. Pharmacol* 2006; 2: 309-315.
- [70] I.A. Al-Mofleh, A.A. Alhaider, J.S. Mossa, M.O. Al-Sohaibani, S. Qureshi and S. Rafatullah. Antigastric Ulcer Studies on 'Saffron' *Crocus sativus* L. in Rats. *Pakistan Journal of Biological Sciences* 2006;9 : 1009-1013 .
- [71] I.A. Al-Mofleh, A.A. Alhaider, J.S. Mossa, M.O. Al-Sohaibani, S. Rafatullah and S. Qureshi. Protection of gastric mucosal damage by *Coriandrum sativum* L. pretreatment in Wistar albino rats. *Environmental Toxicology and Pharmacology* 2006; 22 : 64-69.
- [72] I.A. Al-Mofleh, A.A. Alhaider, J.S. Mossa, M.O. Al-Sohaibani, S. Qureshi and S. Rafatullah. Antisecretagogue, Antiulcer and Cytoprotective Effects of 'Peppermint' *Mentha piperita* L. In *Laboratory Animals. J. Med. Sci* 2006;6 : 930-936.
- [73] Al Mofleh IA, Alhaider AA, Mossa JS, Al-Soohaibani MO, Rafatullah S. Aqueous suspension of anise "*Pimpinella anisum*" protects rats against chemically induced gastric ulcers. *World J Gastroenterol* 2007 ;13 : 1112-1118.
- [74] Kushima H, Nishijima CM, Rodrigues CM, Rinaldo D, Sassá MF, Bauab TM, Stasi LC, Carlos IZ, Brito AR, Vilegas W, Hiruma-Lima CA. *Davilla elliptica* and *Davilla*

- nitida: gastroprotective, anti-inflammatory immunomodulatory and anti-*Helicobacter pylori* action. *J Ethnopharmacol* 2009;123 : 430-438.
- [75] Lima ZP, Calvo TR, Silva EF, Pellizzon CH, Vilegas W, Brito AR, Bauab TM, Hiruma-Lima CA. Brazilian medicinal plant acts on prostaglandin level and *Helicobacter pylori*. *J Med Food* 2008 ;11: 701-708. [PubMed]
- [76] Gomes Rde C, Bonamin F, Darin DD, Seito LN, Di Stasi LC, Dokkedal AL, Vilegas W, Souza Brito AR, Hiruma-Lima CA. Antioxidative action of methanolic extract and buthanolic fraction of *Vochysia tucanorum* Mart. in the gastroprotection.. *J Ethnopharmacol* 2009; 121 : 466-471.
- [77] Suleyman H, Albayrak A, Bilici M, Cadirci E, Halici Z. Different mechanisms in formation and prevention of indomethacin-induced gastric ulcers. *Inflammation* 2010 ;33: 224-234. [PubMed]
- [78] Peskar BM, Maricic N, Gretzera B, Schuligoi R, Schmassmann A. Role of cyclooxygenase-2 in gastric mucosal defense. *Life Sci.* 2001;69: 2993-3003. [PubMed]
- [79] Johansson C, Bergström S. Prostaglandin and protection of the gastroduodenal mucosa. *Scand J Gastroenterol Suppl* 1982;77: 21-46. [PubMed]
- [80] Konturek SJ. Mechanisms of gastroprotection. *Scand J Gastroenterol Suppl* 1990;174: 15-28. [PubMed]
- [81] Falalyeyeva TM, Samonina GE, Beregovaya TV, Andreeva LA, Dvorshchenko EA. Effect of glyprolines PGP, GP, and PG on homeostasis of gastric mucosa in rats with experimental ethanol-induced gastric ulcers. *Bull Exp Biol Med* 2010 ;149 : 699-701. [PubMed]
- [82] Konturek SJ, Konturek PC, Brzozowski T. Prostaglandins and ulcer healing. *J Physiol Pharmacol* 2005;56 Suppl 5: 5-31. [PubMed]
- [83] Gookin JL, Galanko JA, Blikslager AT, Argenzio RA. PG-mediated closure of paracellular pathway and not restitution is the primary determinant of barrier recovery in acutely injured porcine ileum. *Am J Physiol Gastrointest Liver Physiol* 2003;285: G967-G979. [PubMed]
- [84] Hatazawa R, Ohno R, Tanigami M, Tanaka A, Takeuchi K. Roles of endogenous prostaglandins and cyclooxygenase isozymes in healing of indomethacin-induced small intestinal lesions in rats. *J Pharmacol Exp Ther* 2006;318: 691-699. [PubMed]
- [85] Konturek PC, Brzozowski T, Konturek SJ, Taut A, Kwiecien S, Pajdo R, Sliwowski Z, Hahn EG. Bacterial lipopolysaccharide protects gastric mucosa against acute injury in rats by activation of genes for cyclooxygenases and endogenous prostaglandins. *Digestion* 1998;59: 284-297. [PubMed]
- [86] Takayama C, de-Faria FM, Almeida AC, Valim-Araújo DD, Rehen CS, Dunder RJ, Socca EA, Manzo LP, Rozza AL, Salvador MJ, Pellizzon CH, Hiruma-Lima CA, Luiz-Ferreira A, Souza-Brito AR. Gastroprotective and ulcer healing effects of essential oil from *Hyptis spicigera* Lam. (Lamiaceae). *J Ethnopharmacol* 2011 135: 147-55. [PubMed]
- [87] Mehrabani D, Rezaee A, Azarpira N, Fattahi MR, Amini M, Tanideh N, Panjehshahin MR, Saberi-Firouzi M. The healing effects of *Teucrium polium* in the repair of indomethacin-induced gastric ulcer in rats. *Saudi Med J* 2009;30: 494-499. [PubMed]

- [88] Komoike Y, Nakashima M, Nakagiri A, Takeuchi K. Prostaglandin E receptor EPI subtype but not prostacyclin IP receptor involved in mucosal blood flow response of mouse stomachs following barrier disruption. *Digestion* 2003;67: 186-194. [PubMed]
- [89] Satyanarayana MN. Capsaicin and gastric ulcers. *Crit Rev Food Sci Nutr* 2006;46: 275-328. [PubMed]
- [90] Rozza AL, Moraes Tde M, Kushima H, Tanimoto A, Marques MO, Bauab TM, Hiruma-Lima CA, Pellizzon CH. Gastroprotective mechanisms of Citrus lemon (Rutaceae) essential oil and its majority compounds limonene and β -pinene: involvement of heat-shock protein-70, vasoactive intestinal peptide, glutathione, sulfhydryl compounds, nitric oxide and prostaglandin E₂. *Chem Biol Interact* 2011 ;189: 82-89. [PubMed]
- [91] Maity B, Banerjee D, Bandyopadhyay SK, Chattopadhyay S. *Myristica malabarica* heals stomach ulceration by increasing prostaglandin synthesis and angiogenesis. *Planta Med* 2008 ;74: 1774-1778. [PubMed]
- [92] El-Abhar HS. Coenzyme Q10: a novel gastroprotective effect via modulation of vascular permeability, prostaglandin E , nitric oxide and redox status in indomethacin-induced gastric ulcer model. *Eur J Pharmacol* 2010; 649: 314-319. [PubMed]
- [93] Tarnawski AS, Jones MK. The role of epidermal growth factor (EGF) and its receptor in mucosal protection, adaptation to injury, and ulcer healing: involvement of EGF-R signal transduction pathways. *J Clin Gastroenterol* 1998;27 Suppl 1: S12-20. [PubMed]
- [94] Pai R, Tarnawski A. Signal transduction cascades triggered by EGF receptor activation: relevance to gastric injury repair and ulcer healing. *Dig Dis Sci.* 1998 ;43(9 Suppl): 14S-22S. [PubMed]
- [95] Konturek PC, Brzozowski T, Konturek SJ, Ernst H, Drozdowicz D, Pajdo R, Hahn EG. Expression of epidermal growth factor and transforming growth factor alpha during ulcer healing. Time sequence study. *Scand J Gastroenterol* 1997;32: 6-15. [PubMed]
- [96] Jones MK, Tomikawa M, Mohajer B, Tarnawski AS. Gastrointestinal mucosal regeneration: role of growth factors. *Front Biosci* 1999 ;4: D303-309. [PubMed]
- [97] Sasaki E, Tominaga K, Watanabe T, Fujiwara Y, Oshitani N, Matsumoto T, Higuchi K, Tarnawski AS, Arakawa T. COX-2 is essential for EGF induction of cell proliferation in gastric RGM1 cells. *Dig Dis Sci* 2003;48: 2257-2262. [PubMed]
- [98] Milani S, Calabrò A. Role of growth factors and their receptors in gastric ulcer healing. *Microsc Res Tech* 2001; 53 : 360-371. [PubMed]
- [99] Szabo S, Vincze A. Growth factors in ulcer healing: lessons from recent studies. *J Physiol Paris* 2000 ;94 : 77-81. [PubMed]
- [100] Konturek PC, Konturek SJ, Brzozowski T, Ernst H. Epidermal growth factor and transforming growth factor-alpha: role in protection and healing of gastric mucosal lesions. *Eur J Gastroenterol Hepatol* 1995 ;7: 933-937. [PubMed]
- [101] Watanabe S, Hirose M, Wang XE, Kobayashi O, Nagahara A, Murai T, Iwazaki R, Miwa H, Miyazaki A, Sato N. Epithelial-mesenchymal interaction in gastric mucosal restoration. *J Gastroenterol* 2000;35 Suppl 12:65-68 [PubMed] .

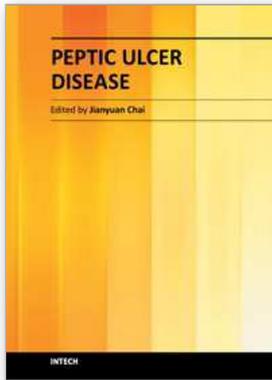
- [102] Tétreault MP, Chailier P, Rivard N, Ménard D. Differential growth factor induction and modulation of human gastric epithelial regeneration. *Exp Cell Res* 2005 ;306 : 285-297 [PubMed].
- [103] Tétreault MP, Chailier P, Beaulieu JF, Rivard N, Ménard D. Epidermal growth factor receptor-dependent PI3K-activation promotes restitution of wounded human gastric epithelial monolayers. *J Cell Physiol* 2008 ;214 : 545-457. [PubMed]
- [104] Ma L, Wang WP, Chow JY, Yuen ST, Cho CH. Reduction of EGF is associated with the delay of ulcer healing by cigarette smoking. *Am J Physiol Gastrointest Liver Physiol* 2000 ;278 : G10-17.
- [105] Cao MB, Dong L, Chang XM, Zou BC, Qin B. Effect of Mexican tea herb and pilular adina herb on concrescence of gastric mucosa in experimental gastric ulcer rats. *Chin J Integr Med* 2007;13: 132-136. [PubMed]
- [106] Ma L, Chow JY, Wong BC, Cho CH. Role of capsaicin sensory nerves and EGF in the healing of gastric ulcer in rats. *Life Sci*. 2000;66: PL213-PL220. [PubMed]
- [107] Wang B, Zhao HY, Zhou L, Wang YF, Cao J. Effect of Kuyangping on expressions of EGF and EGFR mRNA in gastric mucosa in rats with experimental gastric ulcer. *Beijing Zhongyiyao Daxue Xuebao* 2008;31: Abstract.
- [108] Zheng XG, Zhang JJ, Huang YC. [Study on the effect of weitongning on epidermal growth factor and nitric oxide contents in tissue of stomach of rats with gastric ulcer] *Zhongguo Zhongxiyi Jiehe Zazhi* 2004;24: 549-551. [PubMed]
- [109] Meng H, Guo J, Sun JY, Pei JM, Wang YM, Zhu MZ, Huang C. Angiogenic effects of the extracts from Chinese herbs: Angelica and Chuanxiong. *Am J Chin Med* 2008;36: 541-554. [PubMed]
- [110] Ye YN, Koo MW, Li Y, Matsui H, Cho CH. Angelica sinensis modulates migration and proliferation of gastric epithelial cells. *Life Sci* 2001;68: 961-968. [PubMed]
- [111] Debashish Banerjee,1 Biswanath Maity,1 Subrata K Nag,1 Sandip K Bandyopadhyay,1 and Subrata Chattopadhyay 2 Healing Potential of against indomethacin-induced gastric ulceration: a mechanistic exploration. *BMC Complement Altern Med* 2008; 8: 3. [PubMed]
- [112] Banerjee D, Bauri AK, Guha RK, Bandyopadhyay SK, Chattopadhyay S. Healing properties of malabaricone B and malabaricone C, against indomethacin-induced gastric ulceration and mechanism of action. *Eur J Pharmacol* 2008;578: 300-312. [PubMed]
- [113] Banerjee D, Maity B, Bandivdeker AH, Bandyopadhyay SK, Chattopadhyay S. Angiogenic and cell proliferating action of the natural diarylnonanoids, malabaricone B and malabaricone C during healing of indomethacin-induced gastric ulceration. *Pharm Res* 2008 ;25: 1601-1609. [PubMed]
- [114] Seo JY, Yu JH, Lim JW, Mukaida N, Kim H. Nitric oxide-induced IL-8 expression is mediated by NF-kappaB and AP-1 in gastric epithelial AGS cells. *J Physiol Pharmacol* 2009 ;60 Suppl 7: 101-106.
- [115] Li Ma, John L. Wallace. Endothelial nitric oxide synthase modulates gastric ulcer healing in rats. *Am J Physiol Gastrointest Liver Physiol* 2000 279: G341-G346.
- [116] Wang L, Zhou Y, Peng J, Zhang Z, Jiang DJ, Li YJ. Role of endogenous nitric oxide synthase inhibitor in gastric mucosal injury. *Clin Biochem* 2007;40: 615-622. [PubMed]

- [117] Reyes-Trejo B, Sánchez-Mendoza ME, Becerra-García AA, Cedillo-Portugal E, Castillo-Henkel C, Arrieta J. Bioassay-guided isolation of an anti-ulcer diterpenoid from *Croton reflexifolius*: role of nitric oxide, prostaglandins and sulfhydryls. *J Pharm Pharmacol* 2008;60: 931-936. [PubMed]
- [118] Zayachkivska OS, Konturek SJ, Drozdowicz D, Brzozowski T, Gzhegotsky MR. Influence of plant-originated gastroprotective and antiulcer substances on gastric mucosal repair. *Fiziol Zh* 2004;50: 118-127. [PubMed]
- [119] Szabo S. Experimental basis for a role for sulfhydryls and dopamine in ulcerogenesis: a primer for cytoprotection—organoprotection. *Klin Wochenschr* 1986;64 Suppl 7: 116-122. [PubMed]
- [120] Chen SH, Liang YC, Chao JC, Tsai LH, Chang CC, Wang CC, Pan S. Protective effects of *Ginkgo biloba* extract on the ethanol-induced gastric ulcer in rats. *World J Gastroenterol* 2005;11: 3746-3750. [PubMed]
- [121] Chatterjee A, Chattopadhyay S, Bandyopadhyay SK. Biphasic Effect of *Phyllanthus emblica* L. Extract on NSAID-Induced Ulcer: An Antioxidative Trail Weaved with Immunomodulatory Effect. *Evid Based Complement Alternat Med* 2011;2011: 146808.
- [122] M. A. Garcia-Gonzalez, A. Lanasa, S. Santolaria, J. B. A. Crusius, M. T. Serrano, A. S. Peña. The polymorphic IL-1B and IL-1RN genes in the aetiopathogenesis of peptic ulcer. *Clinical & Experimental Immunology* 2001; 125 : 368-375.
- [123] Toshio Watanabe, Kazuhide Higuchi, Masaki Hamaguchi, Masatsugu Shiba, Kazunari Tominaga, Yasuhiro Fujiwara, Takayuki Matsumoto, and Tetsuo Arakawa. Monocyte chemoattractant protein-1 regulates leukocyte recruitment during gastric ulcer recurrence induced by tumor necrosis factor- α . *Am J Physiol Gastrointest Liver Physiol* 2004 287: G919-G928
- [124] R Rad, A Dossumbekova, B Neu, R Lang, S Bauer, D Saur, M Gerhard, C Prinz. Cytokine gene polymorphisms influence mucosal cytokine expression, gastric inflammation, and host specific colonisation during *Helicobacter pylori* infection. *Gut* 2004;53:1082-1089.
- [125] T Watanabe, K Higuchi, K Tominaga, Y Fujiwara, T Arakawa. Acid regulates inflammatory response in a rat model of induction of gastric ulcer recurrence by interleukin 1 β . *Gut* 2001;48: 774-781.
- [126] C. Lindholm, M. Quiding-Järbrink, H. Lönnroth, A. Hamlet, and A.-M. Svennerholm. Local Cytokine Response in *Helicobacter pylori*-Infected Subjects. *Infection and Immunity* 1998; 66 : 5964-5971.
- [127] Carlo-F. Zambon, Daniela Basso, Filippo Navaglia, Claudio Belluco, Alessandra Falda, Paola Fogar, Eliana Greco, Nicoletta Gallo, Massimo Rugge, Francesco Di Mario, and Mario Plebani. Pro- and anti-inflammatory cytokines gene polymorphisms and *Helicobacter pylori* infection: interactions influence outcome. *Cytokine* 2005;29: 141-152.
- [128] Augusto AC, Miguel F, Mendonça S, Pedrazzoli J Jr, Gurgueira SA. Oxidative stress expression status associated to *Helicobacter pylori* virulence in gastric diseases. *Clin Biochem* 2007;40 : 615-622.
- [129] Sugimoto M, Furuta T, Shirai N, Nakamura A, Xiao F, Kajimura M, Sugimura H, Hishida A. Different effects of polymorphisms of tumor necrosis factor- α and

- interleukin-1 beta on development of peptic ulcer and gastric cancer. *J Gastroenterol Hepatol* 2007;22 : 51-59.
- [130] Mitsushige Sugimoto, Yoshio Yamaoka, Takahisa Furuta. Influence of interleukin polymorphisms on development of gastric cancer and peptic ulcer. *World J Gastroenterol* 2010; 16 : 1188-1200.
- [131] Mahattanadul S, Nakamura T, Panichayupakaranant P, Phdoongsombut N, Tungsinnunkong K, Bouking P. Comparative antiulcer effect of bisdemethoxycurcumin and curcumin in a gastric ulcer model system. *Phytomedicine* 2009;16 : 342-351.
- [132] Alexa T. Smolinski and James J. Pestka. Modulation of lipopolysaccharide-induced proinflammatory cytokine production in vitro and in vivo by the herbal constituents apigenin (chamomile), ginsenoside Rb1 (ginseng) and parthenolide (feverfew). *Food and Chemical Toxicology* 2003; 41 : 1381-1390.
- [133] Mahady GB, Pendland SL, Yun G, Lu ZZ. Turmeric (*Curcuma longa*) and curcumin inhibit the growth of *Helicobacter pylori*, a group 1 carcinogen. *Anticancer Res* 2002;22: 4179-4181. [PubMed]
- [134] Salem EM, Yar T, Bamosa AO, Al-Quorain A, Yasawy MI, Alsulaiman RM, Randhawa MA Comparative study of *Nigella Sativa* and triple therapy in eradication of *Helicobacter Pylori* in patients with non-ulcer dyspepsia. *Saudi J Gastroenterol* 2010 ; 6 : 207-214.
- [135] Ali SM, Khan AA, Ahmed I, Musaddiq M, Ahmed KS, Polasa H, Rao LV, Habibullah CM, Sechi LA, Ahmed N. Antimicrobial activities of Eugenol and Cinnamaldehyde against the human gastric pathogen *Helicobacter pylori*. *Ann Clin Microbiol Antimicrob* 2005;4: 20. [PubMed]
- [136] O'Mahony R, Al-Khtheeri H, Weerasekera D, Fernando N, Vaira D, Holton J, Basset C. Bactericidal and anti-adhesive properties of culinary and medicinal plants against *Helicobacter pylori*. *World J Gastroenterol* 2005;11: 7499-7507. [PubMed]
- [137] Shikov AN, Pozharitskaya ON, Makarov VG, Kvetnaya AS. Antibacterial activity of *Chamomilla recutita* oil extract against *Helicobacter pylori*. *Phytother Res* 2008 ;22 :252-253.
- [138] Chun, S.-S., Vattem, D.A., Lin, Y.-T., Shetty K. Phenolic antioxidants from clonal oregano (*Origanum vulgare*) with antimicrobial activity against *Helicobacter pylori*. *Process Biochemistry* 2005;40 : 809-816.
- [139] Zaidi SF, Yamada K, Kadowaki M, Usmanghani K, Sugiyama T. Bactericidal activity of medicinal plants, employed for the treatment of gastrointestinal ailments, against *Helicobacter pylori*. *J Ethnopharmacol* 2009 ;121: 286-291.
- [140] Wang YC, Huang TL Screening of anti-*Helicobacter pylori* herbs deriving from Taiwanese folk medicinal plants. *FEMS Immunol Med Microbiol* 2005 ;43 : 295-300.
- [141] Castillo-Juárez I, González V, Jaime-Aguilar H, Martínez G, Linares E, Bye R, Romero I. Anti-*Helicobacter pylori* activity of plants used in Mexican traditional medicine for gastrointestinal disorders. *J Ethnopharmacol* 2009 ;122 : 402-405.
- [142] Stamatis G, Kyriazopoulos P, Golegou S, Basayiannis A, Skaltsas S, Skaltsa H. In vitro anti-*Helicobacter pylori* activity of Greek herbal medicines. *J Ethnopharmacol* 2003 ;88 : 175-179.
- [143] J.B. Calixto Efficacy, safety, quality control, marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents) *Braz J Med Biol Res* 2000; 33: 179-189.

- [144] Balaji S, Chempakam B. Pharmacokinetics prediction and drugability assessment of diphenylheptanoids from turmeric (*Curcuma longa* L) *Med Chem* 2009;5: 130-138. [PubMed]
- [145] Kandarkar SV, Sawant SS, Ingle AD, Deshpande SS, Maru GB. Subchronic oral hepatotoxicity of turmeric in mice--histopathological and ultrastructural studies. *Indian J Exp Biol* 1998;36: 675-679. [PubMed]
- [146] Shah AH, Qureshi S, Ageel AM. Toxicity studies in mice of ethanol extracts of *Foeniculum vulgare* fruit and *Ruta chalepensis* aerial parts. *J Ethnopharmacol* 1991;34: 167-172. [PubMed]
- [147] Shah AH, Al-Shareef AH, Ageel AM, Qureshi S. Toxicity studies in mice of common spices, *Cinnamomum zeylanicum* bark and *Piper longum* fruits. *Plant Foods Hum Nutr* 1998;52: 231-239. [PubMed]
- [148] Teschke R. Kava hepatotoxicity--a clinical review. *Ann Hepatol* 2010;9: 251-265. [PubMed]
- [149] Nin Chau T, Cheung WI, Ngan T, Lin J, Lee KW, Tat Poon W, Leung VK, Mak T, Tse ML; Hong Kong Herb-Induced Liver Injury Network (HK-HILIN). Causality assessment of herb-induced liver injury using multidisciplinary approach and Roussel Uclaf Causality Assessment Method (RUCAM). *Clin Toxicol (Phila)* 2011;49: 34-39. [PubMed].
- [150] Deshpande SS, Lalitha VS, Ingle AD, Raste AS, Gadre SG, Maru GB. Subchronic oral toxicity of turmeric and ethanolic turmeric extract in female mice and rats. *Toxicol Lett* 1998;95:183-193. [PubMed]
- [151] Daware MB, Mujumdar AM, Ghaskadbi S. Reproductive toxicity of piperine in Swiss albino mice. *Planta Med.* 2000;66:231-236. [PubMed]) De Smet PA. Health risks of herbal remedies. *Drug Saf* 1995;13 : 81-93.
- [152] De Smet PA. Health risks of herbal remedies. *Drug Saf* 1995 ;13: 81-93. [PubMed]
- [153] Markowitz JS, DeVane CL. The emerging recognition of herb-drug interactions with a focus on St. John's wort (*Hypericum perforatum*). *Psychopharmacol Bull* 2001; 35: 53-64. [PubMed]
- [154] Ioannides C. Pharmacokinetic interactions between herbal remedies and medicinal drugs. *Xenobiotica* 2002 ;32 : 451-478. [PubMed]
- [155] Mannel M. Drug interactions with St John's wort: mechanisms and clinical implications. *Drug Saf* 2004;27 : 773-797. [PubMed]
- [156] Borrelli F, Izzo AA. Herb-drug interactions with St John's wort (*Hypericum perforatum*): an update on clinical observations. *AAPS J.* 2009 ;11: 710-27.
- [157] Izzo AA, Ernst E. Interactions between herbal medicines and prescribed drugs: an updated systematic review. *Drugs* 2009;69 : 1777-1798. [PubMed]
- [158] Dasgupta A. Herbal supplements and therapeutic drug monitoring: focus on digoxin immunoassays and interactions with St. John's wort. *Ther Drug Monit* 2008 ;30 : 212-217. [PubMed]
- [159] Dasgupta A, Tso G, Szelei-Stevens K. St. John's wort does not interfere with therapeutic drug monitoring of 12 commonly monitored drugs using immunoassays. *Clin Lab Anal* 2006;20: 62-67. [PubMed]
- [160] Hafner-Blumenstiel V. [Herbal drug-drug interaction and adverse drug reactions]. *Ther Umsch* 2011;68: 54-57. [PubMed].

- [161] Gurley BJ, Gardner SF, Hubbard MA, Williams DK, Gentry WB, Cui Y, Ang CY. Clinical assessment of effects of botanical supplementation on cytochrome P450 phenotypes in the elderly: St John's wort, garlic oil, Panax ginseng and Ginkgo biloba. *Drugs Aging* 2005;22 : 525-539. [PubMed]
- [162] Dugo M, Gatto R, Zagatti R, Gatti P, Cascone C. [Herbal remedies: nephrotoxicity and drug interactions]. *G Ital Nefrol* 2010 ;27 Suppl 52: S5-9. [PubMed]
- [163] Shaw D. Toxicological risks of Chinese herbs. *Planta Med* 2010;76: 2012-2018. [PubMed]
- [164] Cuzzolin L, Francini-Pesenti F, Verlato G, Joppi M, Baldelli P, Benoni G. Use of herbal products among 392 Italian pregnant women: focus on pregnancy outcome. *Pharmacoepidemiol Drug Saf* 2010 ;19 : 1151-1158. [PubMed]
- [165] Westfall RE. Use of anti-emetic herbs in pregnancy: women's choices, and the question of safety and efficacy. *Complement Ther Nurs Midwifery* 2004 ;10 : 30-36.
- [166] Chattopandhyay I, Biswas K, Bandyopadhyay U. Turmeric and curcumin: Biological actions and medicinal applications. *Current Science* 2004; 87: 44-53.



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Peptic ulcer disease is one of the most common chronic infections in human population. Despite centuries of study, it still troubles a lot of people, especially in the third world countries, and it can lead to other more serious complications such as cancers or even to death sometimes. This book is a snapshot of the current view of peptic ulcer disease. It includes 5 sections and 25 chapters contributed by researchers from 15 countries spread out in Africa, Asia, Europe, North America and South America. It covers the causes of the disease, epidemiology, pathophysiology, molecular-cellular mechanisms, clinical care, and alternative medicine. Each chapter provides a unique view. The book is not only for professionals, but also suitable for regular readers at all levels.

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